

BRIEF COMMUNICATION

Carbonic anhydrase, photosynthesis, and seed yield in mustard plants treated with phytohormones

S. HAYAT, A. AHMAD*, M. MOBIN, Q. FARIDUDDIN, and Z.M. AZAM

Plant Physiology Section, Department of Botany, Aligarh Muslim University, Aligarh-202002, India

Abstract

The leaves of 30-d-old plants of *Brassica juncea* Czern & Coss cv. Varuna were sprayed with 10^{-6} M aqueous solutions of indole-3-yl-acetic acid (IAA), gibberellic acid (GA_3), kinetin (KIN), and abscisic acid (ABA) or 10^{-8} M of 28-homobrassinolide (HBR). All the phytohormones, except ABA, improved the vegetative growth and seed yield at harvest, compared with those sprayed with deionised water (control). HBR was most prominent in its effect, generating 32, 30, 36, 70, 25, and 29 % higher values for dry mass, chlorophyll content, carbonic anhydrase (E.C. 4.2.1.1) activity, and net photosynthetic rate in 60-d-old plants, pods per plant, and seed yield at harvest, over the control, respectively. The order of response to various hormones was HBR > GA_3 > IAA > KIN > control > ABA.

Additional key words: abscisic acid; *Brassica juncea* L.; chlorophyll content; gibberellic acid; indole-3-yl-acetic acid; kinetin; net photosynthetic rate; pod and seed number; seed mass and yield.

Various phytohormones exhibit specialised effects on the properties of the membrane and the transport of substances through them, according to their physiological functions. In addition to other functions, auxins affect the proton pump (Aducci *et al.* 1985), gibberellins alter membrane permeability to ions (Crozier and Turnbull 1984), cytokinins favour membrane selectivity by involving calcium (Olah *et al.* 1983), and abscisic acid regulates water movement by modifying K^+/Na^+ selectivity (Knight *et al.* 1995). These and other related factors may have a direct impact on the leaf chlorophyll (Chl) content (Ghorbanli *et al.* 1999, Singh *et al.* 1999). Depending on the plant interaction with the phytohormone, its content increased by auxin application and decreased by GA_3 . Assimilatory pattern modified by phytohormones (Chatterjee *et al.* 1976) may establish a canopy structure most suited for optimal interception of radiant energy favouring higher biological yield (Hayat *et al.* 2000) in mustard. The goal of this experiment was to maximise the biological yield by improving light-harvesting assembly to ensure higher net photosynthetic rate (P_N) through the application of various phytohormones to the plants.

The seeds of *Brassica juncea* Czern & Coss cv. Varuna were obtained from National Seed Corporation Ltd., New Delhi, India. The healthy seeds were surface sterilised with 0.1 % mercuric chloride solution followed by washings with double distilled water and sown in earthen pots (25 cm diameter), lined on its inner surface with polythene sleeves, filled with acid washed sand (Hewitt 1966). Each pot was supplied with 200 cm³ of full nutrient solution (Hoagland and Arnon 1950) on alternate days, up to day 30. Thereafter, the quantity of nutrient solution was increased to 500 cm³. De-ionised water (250 cm³) was also given to each pot, daily. Based on our studies (unpublished), 10^{-6} M aqueous solutions of IAA, GA_3 , KIN, ABA, and 10^{-8} M of HBR were sprayed on 30-d-old plants with the precaution that the phytohormone must not fell in the pot and must stay on the plants. Control plants were sprayed with double distilled water. Each treatment was replicated five times. Dry mass per plant (recorded at 80 °C), chlorophyll (Chl) *a* and *b* contents and Chl *a/b* ratio, carbonic anhydrase (CA) activity, and P_N in the leaves were assessed in 60-d-old plants. The yield characteristics were recorded at harvest. The Chl content was estimated following the

Received 9 October 2000, accepted 11 January 2001.

Fax: +91-571-400528; e-mail: shayat@mailcity.com

Abbreviations: ABA - abscisic acid; CA - carbonic anhydrase; Chl - chlorophyll; DAS - days after sowing; GA_3 - gibberellic acid; HBR - 28-homobrassinolide; IAA - indole-3-yl-acetic acid; KIN - kinetin; P_N - net photosynthetic rate.

Acknowledgements: Authors thank Dr. B.N. Vyas, General Manager, Godrej Agrovet Ltd., Mumbai, India for the generous supply of 28-homobrassinolide.

method of Mackinney (1941). The procedure of Dwivedi and Randhawa (1974) was used for measuring the activity of CA. P_N in intact leaves was measured by LI-6200 portable photosynthesis system (LI-COR, Lincoln,

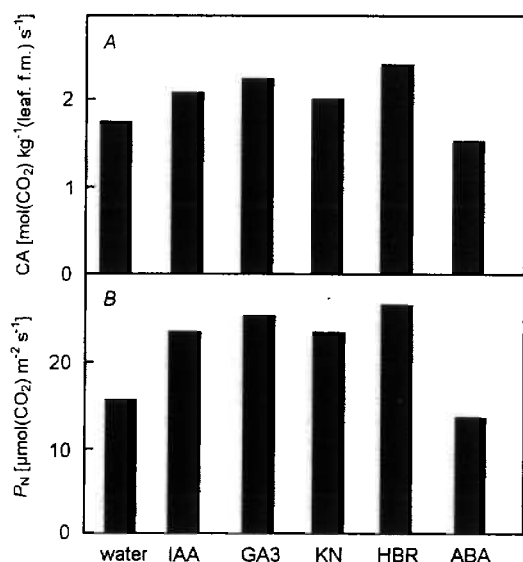


Fig. 1. Effect of phytohormones on leaf carbonic anhydrase (CA) activity (A) and net photosynthetic rate, P_N (B) in 60-d-old mustard plants.

Table 1. Effect of phytohormones on dry mass, yield characteristics, seed yields, and leaf chlorophyll (Chl) content in 60-d-old mustard plants. NS = non-significant. For abbreviations see title page of the paper.

Parameter	Control	IAA	GA ₃	KIN	HBR	ABA	C.D. at 5 %
Dry mass [g plant ⁻¹]	2.25	2.65	2.77	2.50	2.98	1.97	0.15
Pod number [plant ⁻¹]	216.65	250.15	259.15	250.45	271.50	194.15	8.35
Seed number [pod ⁻¹]	12.50	12.53	12.62	12.52	12.60	12.43	NS
100 seed mass [mg]	320.15	321.45	324.15	322.00	323.00	320.45	NS
Seed yield [g plant ⁻¹]	7.24	8.10	8.64	8.25	9.35	6.83	0.38
Chl <i>a</i> [mg kg ⁻¹]	502	635	662	581	755	401	80
Chl <i>b</i> [mg kg ⁻¹]	753	899	952	851	997	623	70
Chl (<i>a</i> + <i>b</i>) [mg kg ⁻¹]	1255	1535	1615	1432	1752	1025	130
Chl <i>a/b</i>	0.67	0.71	0.70	0.69	0.76	0.64	0.07

carboxylase/oxygenase (RuBPCO) and P_N (Davies and Jones 1991, Makheev *et al.* 1992). Similar conclusions may be derived from our observations (Fig. 1B) where all the phytohormones studied, except ABA, enhanced P_N of mustard leaves. HBR was best increasing P_N by 29 % over the control. The higher P_N of the plants receiving phytohormones may be explained by the observed increase in Chl contents (Table 1), higher P_N and lower stomatal resistance (Feierabend 1969, Treharne *et al.* 1970, Hoad *et al.* 1977, Arteca and Dong 1981), and elevated amount of photophosphorylation in isolated chloroplast (Tamas *et al.* 1973).

The activity of CA was increased by the application to the leaves of all the phytohormones, except ABA (Fig.

NE, USA). The results were statistically analysed following the procedures described by Gomez and Gomez (1984).

All the parameters studied on day 60 were significantly enhanced by all the phytohormones, except ABA. HBR proved best, enhancing plant dry mass, leaf Chl *a*, *b*, and (*a*+*b*) contents, Chl *a/b* ratio, CA activity, and P_N by 39.6, 50.3, 32.4, 13.6, 35.8, and 68.8 %, respectively, over the control (Table 1, Fig. 1). IAA and KIN also induced a significant response but the values were equal. Similarly, pod number per plant and seed yield at harvest were significantly enhanced by all the phytohormones studied, except ABA. A maximum increase of 29.1 % more seeds over the control was recorded with HBR. However, the plants sprayed with ABA yielded 5.6 % less seeds.

Higher P_N in the leaves may successfully be attained by the exogenous application of aqueous solution of IAA (Turner and Bidwell 1965, Chatterjee *et al.* 1976), GA₃ (Gale *et al.* 1974, Erkan and Bangerth 1980), KIN/benzyladenine (Treharne *et al.* 1970), or BR/HBR (Braun and Wild 1984, Luo *et al.* 1986, Yang *et al.* 1992, Hayat *et al.* 2000). However, ABA is assigned some important regulatory roles in protecting the plants from stress (Markhart *et al.* 1979, Davies and Jones 1991) but brings about stomata closure resulting in a negative impact on the amount of ribulose-1,5-bisphosphate

1A) which may be because of its *de novo* synthesis by involving transcription and/or translation (Okabe *et al.* 1980). CA is found in abundance in the photosynthesising tissue of both C₃ and C₄ plants and regulates the availability of CO₂ to RuBPCO by catalysing the reversible hydration of CO₂ (Badger and Price 1994). This makes the distribution patterns of CA and RuBPCO comparable to each other (Tsuzuki *et al.* 1985). Moreover, CA should also have some role in electron transport chain (Stemler 1997) and regulation of pH in chloroplasts exposed to rapidly changing irradiance because of its association with thylakoids (Reed and Graham 1981). Antisense expression has been employed to identify the specific role(s) of chloroplast located CA in C₃ plants

(Majeau *et al.* 1994, Price *et al.* 1994). In antisense plants only a 5 % decline in P_N was found by a 1 % decrease in CA activity (Williams *et al.* 1996). In contrast, an inhibition of CA activity in ethoxymolamide infiltrated C_3 plant leaf pieces resulted in 80-90 % inhibition of P_N at low CO_2 concentrations. Thus some more essential role could be assigned to CA than that noted in antisense studies (Badger and Pfanz 1995).

The leaves of the plants, treated with HBR, GA_3 , IAA, and KIN, photosynthesised at a faster rate than the

control (Fig. 1B) and were possibly withheld for a longer duration because of delayed senescence (Menon and Srivastava 1984) and abscission (Iwahori *et al.* 1990). Thus availability of photosynthates in larger quantities for a longer duration may have contributed favourably in improving pod bearing capacity of the plants leading to an increase in seed yield at harvest (Table 1). The order of response to various phytohormones is similar for all the parameters studied (HBR > GA_3 > IAA > KIN > control > ABA).

References

- Aducci, P., Ballio, A., Marra, M.: Incubation of corn coleoptiles with auxin enhances in vitro fusicoccin binding. – *Planta* **167**: 129-132, 1985.
- Arteca, R.N., Dong, C.-N.: Increased photosynthetic rates following gibberellic acid treatments to the roots of tomato plants. – *Photosynth. Res.* **2**: 243-249, 1981.
- Badger, M.R., Pfanz, H.: Effect of carbonic anhydrase inhibition on photosynthesis by leaf pieces of C_3 and C_4 plants. – *Aust. J. Plant Physiol.* **22**: 45-49, 1995.
- Badger, M.R., Price, G.D.: The role of carbonic anhydrase in photosynthesis. – *Annu. Rev. Plant Physiol. Plant mol. Biol.* **45**: 369-392, 1994.
- Braun, P., Wild, A.: The influence of brassinosteroid on growth and parameters of photosynthesis of wheat and mustard plants. – *J. Plant Physiol.* **116**: 189-196, 1984.
- Chatterjee, A., Mandal, R.K., Sircar, S.M.: Effects of growth substances on productivity, photosynthesis and translocation of rice varieties. – *Indian J. Plant Physiol.* **19**: 131-138, 1976.
- Crozier, A., Turnbull, C.: Gibberellins: Biochemistry and action in extension growth. – *What's new Plant Physiol.* **15**: 9-12, 1984.
- Davies, W.J., Jones, H.G.: Absciscic Acid: Physiology and Biochemistry. – Bios Scientific Publ., Oxford 1991.
- Dwivedi, R.S., Randhawa, N.S.: Evaluation of a rapid test for the hidden hunger of zinc in plants. – *Plant Soil* **40**: 445-451, 1974.
- Erkan, Z., Bangerth, F.: Untersuchungen über den Einfluss von Phytohormonen und Wachstumsregulatoren auf den Wasserverbrauch, das Stomataverhalten und die Photosynthese von Paprika- und Tomatenpflanzen. – *Angew. Bot.* **54**: 207-220, 1980.
- Feierabend, J.: Der Einfluss von Cytokininen auf die Bildung von Photosyntheseenzymen in Roggenkeimlingen. – *Planta* **84**: 11-29, 1969.
- Gale, M.D., Edrich, J., Lupton, F.G.H.: Photosynthetic rates and the effects of applied gibberellins in some dwarf, semi-dwarf and tall wheat varieties (*Triticum aestivum*). – *J. agr. Sci.* **83**: 43-46, 1974.
- Ghorbanli, M., Kaveh, S.H., Sepehr, M.F.: Effects of cadmium and gibberellin on growth and photosynthesis of *Glycine max*. – *Photosynthetica* **37**: 627-631, 1999.
- Gomez, K.A., Gomez, A.A.: Statistical Procedures for Agricultural Research. – John Wiley & Sons, New York 1984.
- Hayat, S., Ahmad, A., Mobin, M., Hussain, A., Fariduddin, Q.: Photosynthetic rate, growth and yield of mustard plants sprayed with 28-homobrassinolide. – *Photosynthetica* **38**: 469-471, 2000.
- Hewitt, E.J.: Sand and Water Culture Methods Used in the Study of Plant Nutrition. – Commonwealth Agricultural Bureaux, Farnham Royal 1966.
- Hoad, G.V., Loveys, B.R., Skenek, G.M.: The effect of fruit removal on cytokinins and gibberellin like substance. – *Planta* **136**: 25-30, 1977.
- Hoagland, D.R., Arnon, D.I.: The water culture method for growing plants without soil. – *Univ. Calif. Exp. Stat. Circ.* **347**, 1950.
- Iwahori, S., Tomonaga, S., Higuchi, S.: Retardation of abscission of citrus leaf and fruitlet explant by brassinolide. – *Plant Growth Regul.* **9**: 119-125, 1990.
- Knight, D.C., Sehgal, A., Atwal, K., Wallace, J.C., Cove, D.J., Coates, D., Quatrano, R.S., Bahadur, S., Stockley, P.G., Cuming, A.C.: Molecular responses to abscisic acid and stress are conserved between moss and cereals. – *Plant Cell* **7**: 499-506, 1995.
- Luo, B.S., Kumura, A., Ishji, R., Wada, Y.: The effect of brassinolide treatment on growth and development of wheat. – *J. Crop Sci.* **55**: 29-37, 1986.
- Mackinney, G.: Absorption of light by chlorophyll solutions. – *J. biol. Chem.* **140**: 315-322, 1941.
- Majeau, N., Arnoldo, M.A., Coleman, J.R.: Modification of carbonic anhydrase activity by antisense and over expression constructs in transgenic tobacco. – *Plant mol. Biol.* **25**: 377-385, 1994.
- Makheev, A.V., Krendeleva, T.E., Mokronosov, A.T.: Photosynthesis and abscisic acid. – *Soviet Plant Physiol.* **39**: 118-126, 1992.
- Markhart, A.H., Fiscus, E.L., Naylor, A.W., Kramer, P.J.: Effect of abscisic acid on root hydraulic conductivity. – *Plant Physiol.* **64**: 611-614, 1979.
- Menon, K.K.G., Srivastava, H.C.: Increasing plant productivity through improved photosynthesis. – *Proc. Indian Acad. Sci. (Plant Sci.)* **93**: 359-378, 1984.
- Okabe, K., Lindlar, A., Tsuzuki, M., Miyachi, S.: Effects of carbonic anhydrase on ribulose 1,5-bisphosphate carboxylase and oxygenase. – *FEBS Lett.* **114**: 142-144, 1980.
- Olah, Z., Berczi, A., Erdei, L.: Benzylaminopurine-induced coupling between calmodulin and Ca-ATPase in wheat root microsomal membranes. – *FEBS Lett.* **154**: 395-399, 1983.
- Price, G.D., Caemmerer, S. von, Evans, J.R., Yu, J.-W., Lloyd, J., Oja, V., Kell, P., Harrison, K., Gallagher, A., Badger, M.R.: Specific reduction of chloroplast carbonic anhydrase activity by antisense RNA in transgenic tobacco has a minor effect on photosynthetic CO_2 assimilation. – *Planta* **193**: 331-340, 1994.

- Reed, M.L., Graham, D.: Carbonic anhydrase in plants, distribution, properties and possible physiological roles. – *Phytochemistry* **7**: 47-94, 1981.
- Singh, P., Srivastava, N.K., Mishra, A., Sharma, S.: Influence of etherel and gibberellic acid on carbon metabolism, growth, and essential oil accumulation in spearmint (*Mentha spicata*). – *Photosynthetica* **36**: 509-517, 1999.
- Stemler, A.J.: The case for chloroplast thylakoid carbonic anhydrase. – *Physiol. Plant.* **99**: 348-353, 1997.
- Tamas, I.A., Schwartz, J.W., Breithaupt, B.J., Hagin, J.M., Arnold, P.H.: Effect of indole acetic acid on photosynthetic reactions in isolated chloroplast. – *Proc. Eighth International Conference on Plant Growth Substances*. Pp. 1159-1168. 1973.
- Treharne, K.J., Stoddart, J.L., Pughe, J., Paranjothy, K., Wareing, P.F.: Effects of gibberellin and cytokinins on the activity of photosynthetic enzymes and plastid ribosomal RNA synthesis in *Phaseolus vulgaris* L. – *Nature* **228**: 129-131, 1970.
- Tsuzuki, M., Miyachi, S., Edward, G.E.: Localization of carbonic anhydrase in mesophyll cells of terrestrial C₃ plants in relation to CO₂ assimilation. – *Plant Cell Physiol.* **26**: 881-891, 1985.
- Turner, W.B., Bidwell, R.G.S.: Rates of photosynthesis in attached and detached bean leaves and the effect of spraying with indole-3-acetic acid. – *Plant Physiol.* **40**: 446-451, 1965.
- Williams, T.G., Flanagan, L.B., Coleman, J.R.: Photosynthetic gas exchange and discrimination against ¹³CO₂ and C¹⁸O¹⁶O tobacco plants modified by an antisense construct to have low chloroplastic carbonic anhydrase. – *Plant Physiol.* **112**: 319-326, 1996.
- Yang, Z.S., Shi, G.A., Jin, J.H.: Effects of epibrassinolide, a growth promoting steroidal lactone. I. Activity in selected bioassays. – *Physiol. Plant.* **53**: 445-452, 1992.