

## BRIEF COMMUNICATION

**Assessment of photosynthetic potential of indoor plants under cold stress**

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**Abstract**

Photosynthetic parameters including net photosynthetic rate ( $P_N$ ), transpiration rate ( $E$ ), water-use efficiency (WUE), and stomatal conductance ( $g_s$ ) were studied in indoor C<sub>3</sub> plants *Philodendron domesticum* (*Pd*), *Dracaena fragans* (*Df*), *Peperomia obtusifolia* (*Po*), *Chlorophytum comosum* (*Cc*), and in a CAM plant, *Sansevieria trifasciata* (*St*), exposed to various low temperatures (0, 5, 10, 15, 20, and 25°C). All studied plants survived up to 0°C, but only *St* and *Cc* endured, while other plants wilted, when the temperature increased back to room temperature (25°C). The  $P_N$  declined rapidly with the decrease of temperature in all studied plants. *St* showed the maximum  $P_N$  of 11.9  $\mu\text{mol m}^{-2} \text{s}^{-1}$  at 25°C followed by *Cc*, *Po*, *Pd*, and *Df*.  $E$  also followed a trend almost similar to that of  $P_N$ . *St* showed minimum  $E$  (0.1  $\text{mmol m}^{-2} \text{s}^{-1}$ ) as compared to other studied C<sub>3</sub> plants at 25°C. The  $E$  decreased up to ~4-fold at 5 and 0°C. Furthermore, a considerable decline in WUE was observed under cold stress in all C<sub>3</sub> plants, while *St* showed maximum WUE. Similarly, the  $g_s$  also declined gradually with the decrease in the temperature in all plants. Among C<sub>3</sub> plants, *Pd* and *Po* showed the maximum  $g_s$  of 0.07  $\text{mol m}^{-2} \text{s}^{-1}$  at 25°C followed by *Df* and *Cc*. However, *St* showed the minimum  $g_s$  that further decreased up to ~4-fold at 0°C. In addition, the content of photosynthetic pigments [chlorophyll *a*, *b*, (*a+b*), and carotenoids] was varying in all studied plants at 0°C. Our findings clearly indicated the best photosynthetic potential of *St* compared to other studied plants. This species might be recommended for improving air quality in high-altitude closed environments.

*Additional key words:* house plants; low-temperature stress; net photosynthetic rate; photosynthetic pigment; stomatal conductance; transpiration rate, water-use efficiency.

Considerable parts of human populations, including armed forces, are deployed in inaccessible snow-bound/high-altitude areas of strategic importance at country borders. The environmental conditions in such areas are very harsh, adverse, and with extremely low temperatures and a very low oxygen content (hypoxia) that creates a lot of physiological and psychological stresses affecting their health and performance (West 1995). Therefore, only oxygen enrichment can counteract the effects of hypoxia under such conditions, since plant produce oxygen as a byproduct of photosynthesis (Wood 2003). Thus, by

placing indoor air-purifier plants could enrich the oxygen content in closed environments at high-altitude (Wolverton *et al.* 1984). It does not increase only an aesthetic value inside such an environment but it can be also used to improve the air quality together with removing carbon dioxide and other harmful gasses (Papinchak *et al.* 2009).

There are several air-purifying indoor C<sub>3</sub> plants, namely, *Philodendron domesticum* (*Pd*), *Dracaena fragans* (*Df*), *Peperomia obtusifolia* (*Po*), *Chlorophytum comosum*

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*Abbreviations:* ANOVA – analysis of variance; Car – carotenoids; *Cc* – *Chlorophytum comosum*; Chl – chlorophyll; *Df* – *Dracaena fragans*; DMF – N,N-dimethyl formamide;  $E$  – transpiration rate; FM – fresh mass;  $g_s$  – stomatal conductance; LSD – least significant difference; *Pd* – *Philodendron domesticum*;  $P_N$  – net photosynthetic rate; *Po* – *Peperomia obtusifolia*; *St* – *Sansevieria trifasciata*; WUE – water-use efficiency.

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(*Cc*), and CAM plant, *Sansevieria trifasciata* (*St*). They are known to be effective in high oxygen productivity even during night hours by significant reduction of indoor air pollutants in a closed environment (Wolverton *et al.* 1984, Maroni *et al.* 1995, Gupta *et al.* 2015).

Low temperature is one of the most important abiotic factors that may limit photosynthetic performance of plants (Oliveira and Peñuelas 2004, Ashraf and Harris 2013). It causes severe restrictions to plant function and growth. Therefore, plants have adapted different types of photosynthetic mechanisms ( $C_3$ ,  $C_4$ , and CAM) for specific climate zones (Orsenigo *et al.* 1997, Doubnerová and Ryšlavá 2011, Freschi and Mercier 2012). Also, the overall photosynthetic performance of plants depends upon various stomatal or nonstomatal factors under cold stress (Freschi and Mercier 2012, Yamori *et al.* 2014). Therefore, it is not easy to discriminate between the effects of these limitations on overall photosynthetic capacity of plants. However, some studies have shown the effects of cold stress on various photosynthetic parameters including net photosynthetic rate ( $P_N$ ), transpiration rate ( $E$ ), water-use efficiency (WUE), stomatal conductance ( $g_s$ ), photosynthetic pigments content, *etc.* in many plants (Roden *et al.* 1999, Oliveira and Peñuelas 2004).

The present study was aimed to assess the photosynthetic potential of some air-purifying indoor  $C_3$  and CAM plants under low temperature in order to select a best candidate for improving air quality in closed, high-altitude environments including army bunkers.

The selected indoor potted plants were grown up to 30–50 cm at ambient room temperature ( $25 \pm 2^\circ\text{C}$ ) in laboratory conditions before exposing them to low-temperature stress. After that, various low temperatures (25, 20, 15, 10, 5, and  $0^\circ\text{C}$ ) were set for one week in a decreasing temperature order in a plant growth chamber *LT-105* (Percival Scientific Inc., Perry, Iowa, USA) equipped with cold fluorescent light [ $350 \mu\text{mol}(\text{photon}) \text{m}^{-2} \text{s}^{-1}$ ] with a photoperiod of 12/12 h (day/night) at 500 ppm  $\text{CO}_2$  and relative humidity of  $70 \pm 2\%$ . All photosynthetic parameters ( $P_N$ ,  $E$ , WUE, and  $g_s$ ) were measured after 1-week adaptation of the plants under respective low temperatures by using a portable leaf chamber analyser (*LCA-4*, ADC Bio Scientific Ltd., Hoddesdon, UK).  $P_N$ ,  $E$ , and  $g_s$  were measured on uppermost, fully expanded, terminal leaflets (abaxial surface) at 11:00 h. The same plants were brought to ambient room temperature ( $25 \pm 2^\circ\text{C}$ ) inside laboratory to overcome the effect of previous cold stress, before exposing them to next low-temperature stress. WUE was calculated as the ratio of the  $P_N$  to  $E$ . Chl *a*, Chl *b*, and total Chl (*a+b*) and total carotenoids (Car) contents were determined from fresh leaf samples (FM). Fresh leaf discs were placed in a test tube containing 10 ml of N,N-dimethyl formamide (DMF) and stored for 24 h at  $4^\circ\text{C}$ . The absorbance of the coloured supernatant was read

at 480, 647, and 666 nm by a monochromator-based multimode detector (BioTek, *Snergy 2*, USA) with DMF as a blank. The contents of Chl *a*, *b*, and total Chl (*a+b*) were calculated according to the method of Moran and Porath (1980).

All measurements were carried out in triplicates by using three independent plants for each treatment. *CropStat* program developed at IRRI, Philippines was used for analysis of variance (*ANOVA*) of experiments. The treatment means were compared by least significant difference (LSD) test at a significance level of  $P \leq 0.05$ .

Each plant species has its unique temperature requirements; thus, what is an optimum for one plant species it might be stressful for another one. In general, photosynthesis can function without any harm between 0 and  $30^\circ\text{C}$  in cold-adapted plants that are active in the winter (Ashraf and Harris 2013). Most plant species are able to acclimate to changes in growth temperature by modifying the photosynthetic apparatus in a manner that helps in their survival under low-temperature stress conditions (Biswal *et al.* 2011).

In our case, all studied plants survived morphologically up to  $0^\circ\text{C}$ , but when the temperature increased later to room temperature of  $25^\circ\text{C}$ , only *St* and *Cc* were able to recover from cold stress ( $0^\circ\text{C}$  for 1 week), while the rest of the plants wilted due to permanent cold injury. The optimum temperature for photosynthesis differs depending on plants species and differs also in dependence on growth and measurement temperatures even in a single plant species (Yamori *et al.* 2010). Among studied plants, *St* showed the best survival potential (Fig. 1S, *supplementary material available online*). Most of the CAM plants show very species-specific behaviour in regard to photosynthetic gas exchange and many of them can switch between  $C_3$  and CAM photosynthesis depending on various environmental conditions including low-temperature stress. However, the selected CAM plant, *St*, is obligatory in its nature since it cannot revert into the  $C_3$  phenotype, when the environmental stress is eliminated.

Under low-temperature stress,  $P_N$  declined rapidly with the decrease in the temperature in all plants (Fig. 1A). Among all plants, *St* showed maximum  $P_N$  of  $11.9 \mu\text{mol} \text{m}^{-2} \text{s}^{-1}$  followed by *Cc*, *Po*, *Pd*, and *Df* with  $P_N$  value of  $8.8$ ,  $7.4$ ,  $7.3$ , and  $4.3 \mu\text{mol} \text{m}^{-2} \text{s}^{-1}$ , respectively, at  $25^\circ\text{C}$  (Fig. 1A). The  $P_N$  was  $\sim 3$ -fold higher in *St* as compared to *Cc*. Also, *St* plant showed minimum  $7.5$  and  $15.5$ -fold decrease in  $P_N$  at  $5$  and  $0^\circ\text{C}$ , respectively, as compared to other studied plants. However, the maximum  $\sim 390$  and  $\sim 33$ -fold decrease in  $P_N$  was observed in *Po* and *Cc* plants, respectively, at  $0^\circ\text{C}$ . Reduction in  $P_N$  due to cold stress may be attributed to impairment in photosynthetic pigment biosynthesis or destruction which leads to impairment in the electron transport (Krivosheeva *et al.* 1996). In our case, the  $P_N$  also declined significantly with a decrease in temperature in all studied plants, which was

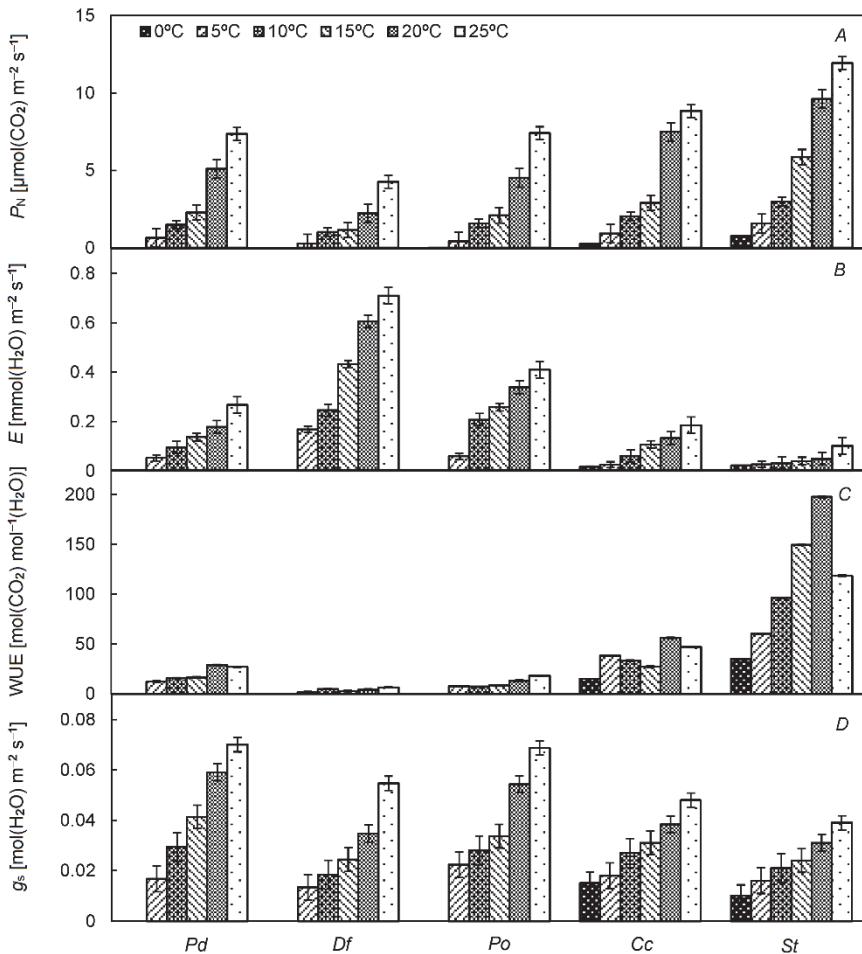


Fig. 1. Effect of various low temperatures (0, 5, 10, 15, 20, and 25°C) on (A) net photosynthetic rate ( $P_N$ ), (B) transpiration rate ( $E$ ), (C) water-use efficiency (WUE), and (D) stomatal conductance ( $g_s$ ) in air-purifying indoor  $C_3$  plants: *Philodendron domesticum* (*Pd*), *Dracaena fragrans* (*Df*), *Peperomia obtusifolia* (*Po*), *Chlorophytum comosum* (*Cc*), and CAM plant, *Sansevieria trifasciata* (*St*). Bar shown means ( $n = 3$ )  $\pm$  SE are statistically significant ( $P \leq 0.05$ ) according to LSD test.

in accordance with earlier reports (Tewari and Tripathy 1998, Biswal *et al.* 2011). The  $C_3$  plants generally possess a greater ability for temperature acclimation of photosynthesis across a broad temperature range (10–35°C), while CAM plants acclimated day and night photosynthetic process differentially with respect to temperature (Yamori *et al.* 2014). Moreover, in CAM plants the optimum temperature for photosynthesis is lower as compared to  $C_3$  and  $C_4$  plants, since their photosynthetic enzymes are stabilized even at low temperatures (Freschi and Mercier 2012). Also, CAM plants are well adapted for cold, dry, and arid environments and have usually higher  $P_N$  as compared to  $C_3$  plants (Freschi and Mercier 2012, Yamori *et al.* 2014). Thus, higher  $P_N$  in *St* was in agreement with above mentioned.

The  $E$  followed a trend almost similar to that of  $P_N$ . A sharp decrease was observed in  $E$  under low-temperature stress in all studied plants except of *St*. Among  $C_3$  plants, *Df* showed maximum  $E$  ( $0.7 \text{ mmol m}^{-2} \text{s}^{-1}$ ) at 25°C that decreased up to ~4-fold upon subsequent low-temperature stress. The higher  $E$  was observed also in *Po* followed by *Pd* and *Cc*. However, *St* plants showed minimum  $E$  ( $0.1 \text{ mmol m}^{-2} \text{s}^{-1}$ ) as compared to other  $C_3$  plants at 25°C; it further smoothly decreased up to ~4-fold at 5 and 0°C

(Fig. 1B). It is known that CAM plants close their stomata during the day to avoid water losses by transpiration (Freschi and Mercier 2012, Yamori *et al.* 2014). Therefore, there was no surprise that the least  $E$  was observed in *St* as compared to other studied  $C_3$  plants.

WUE is defined as the ratio of carbon gain (dry matter accumulation) to water loss (amount of water transpired during a period of dry matter accumulation) (Sinclair *et al.* 1984). A considerable decline in WUE under low-temperature stress was observed in all studied  $C_3$  plants due to the higher  $E$  as compared to the CAM plant, since their stomata remained opened during the daytime (Freschi and Mercier 2012), while in *St* (being the CAM plant), the decrease was comparatively lesser (Fig. 1C). *St* showed maximum WUE as compared to all studied  $C_3$  plants and thus proved its fitness to survive under low temperature. CAM plants can effectively save metabolic energy and water during harsh environmental conditions as compared to  $C_3$  plants (Freschi and Mercier 2012). Among the  $C_3$  plants, *Cc* plants showed the best WUE followed by *Pd*, *Po*, and *Df*. Plants can achieve high WUE through either high  $P_N$ , low  $E$ , or both. Both processes are at least partially regulated by  $g_s$  (Cowan 1982). When water is limiting, conductance decreases as a result of stomatal

Table 1. Effect of low-temperature stress (0°C for 1 week) on alteration of photosynthetic pigments [chlorophyll (Chl) *a*, Chl *b*, total Chl (*a+b*) and carotenoids (Car)] in air-purifying indoor C<sub>3</sub> plants, such as *Philodendron domesticum* (*Pd*), *Dracaena fragans* (*Df*), *Peperomia obtusifolia* (*Po*), *Chlorophytum comosum* (*Cc*) and CAM plant, *Sansevieria trifasciata* (*St*). Means (*n* = 3) ± SE value of the treatments are statistically significant (*P* ≤ 0.05) according to LSD test. FM – fresh mass.

Parameter	Chl <i>a</i> [ $\mu\text{g g}^{-1}$ (FM)]		Chl <i>b</i> [ $\mu\text{g g}^{-1}$ (FM)]		Total Chl ( <i>a+b</i> ) [ $\mu\text{g g}^{-1}$ (FM)]		Car [ $\mu\text{g g}^{-1}$ (FM)]	
Temperature	25°C	0°C	25°C	0°C	25°C	0°C	25°C	0°C
<i>Pd</i>	0.14	0.08	0.07	0.04	0.22	0.13	1.34	1.46
<i>Df</i>	0.02	0.02	0.01	0.01	0.03	0.03	1.05	2.26
<i>Po</i>	0.14	0.41	0.05	0.43	0.19	0.94	1.03	1.75
<i>Cc</i>	0.17	0.05	0.06	0.03	0.24	0.09	1.42	0.60
<i>St</i>	0.26	0.89	0.09	1.07	0.36	2.03	0.70	0.51
SE (M)	0.05	0.08	0.02	0.05	0.07	0.10	0.25	0.04
LSD ( <i>P</i> ≤ 0.05)	0.15	0.28	0.07	0.16	0.22	0.34	0.80	0.13

closure. Stomatal closure has a greater effect on photosynthesis than on transpiration due to the additional resistance associated with diffusion of CO<sub>2</sub> relative to H<sub>2</sub>O in the leaf (Medici *et al.* 2007). Therefore, the increase in WUE caused by stomatal closure resulted in the decrease in the carbon assimilation. However, when water is limited, plants, which use a finite water supply more efficiently, should grow more rapidly (Nobel 1996). CAM plants open their stomata and incorporate CO<sub>2</sub> during night, when evapo-transpiration rates are low; it results in high WUE as compared to C<sub>4</sub> and C<sub>3</sub> plants (Nobel 1996). Therefore, under low-temperature stress, plants with a high WUE should have greater ability to survive than the plants with the low WUE (Sinclair *et al.* 1984); it was in accordance with our results.

The *g<sub>s</sub>* declined gradually with the decrease in the temperature in all studied plants (Fig. 1D). The regulation of leaf *g<sub>s</sub>* is a key phenomenon in plants as it is vital for both prevention of desiccation and CO<sub>2</sub> acquisition (Medici *et al.* 2007). Stomata closure in response to cold stress generally occurs due to decreased leaf turgor and atmospheric pressure ultimately alters photosynthesis and the mesophyll metabolism (Medici *et al.* 2007, Xu *et al.* 2008). Among the C<sub>3</sub> plants, *Pd* and *Po* showed maximum *g<sub>s</sub>* of 0.07 mol m<sup>-2</sup> s<sup>-1</sup> followed by *Df* and *Cc* with *g<sub>s</sub>* value of 0.05 and 0.04 mol m<sup>-2</sup> s<sup>-1</sup>, respectively, at 25°C (Fig. 1D). On the other hand, *St* showed minimum *g<sub>s</sub>* (0.03 mol m<sup>-2</sup> s<sup>-1</sup>), which further decreased up to ~4-fold at 0°C, since the CAM plants close their stomata during daytime (Freschi and Mercier 2012, Yamori *et al.* 2014). Therefore, the higher *g<sub>s</sub>* in the studied C<sub>3</sub> plants as compared to CAM

plants was expected.

The alteration of photosynthetic pigments [Chl *a*, *b*, and total Chl (*a+b*)] content was found in all studied plants at 0°C. The pigment amount was significantly reduced (~0.3 to 0.9-fold) as compared to 25°C in *Pd*, *Df*, and *Cc* plants (Table 1). It is well known that the chloroplast is the main organelle rapidly and deeply affected during cold stress (Ensminger *et al.* 2006). When plants are exposed to low-temperature stress, Chl biosynthesis is inhibited. Therefore, an imbalance in PSII is created after exposure to low temperature because of the alterations in the Chl antenna complexes (Habibi *et al.* 2011). In contrast, in *Po* and *St*, no such decrease in photosynthetic pigments was observed (Table 1). It might suggest a less harmful effect of Chl bleaching and photoinhibition under cold stress (Habibi *et al.* 2011). Similarly, the total carotenoids (Car) content was also found varying in all studied plants at 0°C. In *Pd*, *Df*, and *Po* plants, the Car content slightly increased (~1.1 to 2-fold) at 0°C as compared to 25°C (Table 1). At low temperature, the alterations in the content of total Car showed a tendency to a sharp increase, probably in order to increase photon capture, as a strategy against cold-induced photoinhibition (Habibi *et al.* 2011). However, in *Cc* and *St*, no significant change in the Car content was observed at 0°C (Table 1), which might suggest their better survival potential at 0°C.

Our findings clearly indicated the best photosynthetic potential of *Sansevieria trifasciata* (*St*) compared to other studied plants. Therefore, it may be recommended to be pot-grown in high-altitude closed environments for improving air quality under low temperature.

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