

The effect of CaCl₂ on calcium content, photosynthesis, and chlorophyll fluorescence of tung tree seedlings under drought conditions

Z. LI^{*,**}, X.F. TAN^{*,**,†}, K. LU^{*,**}, Z.M. LIU^{*,**,***}, and L.L. WU^{*,**}

Key Laboratory of Cultivation and Protection for Non-Wood Forest Trees, Ministry of Education,

Central South University of Forestry and Technology, Changsha 410004, China^{}*

Cooperative Innovation Center of Cultivation and Utilization for Non-Wood Forest Trees of Hunan Province,

*Central South University of Forestry and Technology, Changsha 410004, China^{**}*

*Department of Biology, Eastern New Mexico University, Portales, NM 88130, USA^{***}*

Abstract

The study investigated the effects of different CaCl₂ concentrations (2, 5, and 10 mM) on photosynthetic enzymatic activities, photosynthesis, and chlorophyll fluorescence of tung tree seedlings under drought conditions. Plants were sprayed with either CaCl₂ or distilled water until run-off. Irrigation was then withheld to induce drought stress. The strength of drought stress was evaluated by relative leaf water content and soil water content, which was 27.3 and 9.5% on day 0 and day 12, respectively. Drought stress decreased activities of ribulose-1,5-bisphosphate carboxylase/oxygenase and phosphoenolpyruvate carboxylase, chlorophyll (*a+b*) content, net photosynthetic rate, stomatal conductance, transpiration rate, electron transport rate, the maximal quantum yield of PSII photochemistry, and effective quantum yield of PSII in tung tree seedlings. The CaCl₂ pretreatments alleviated the negative effect of drought stress to some degree on all the parameters mentioned above.

Additional key words: calcium chloride; gas exchange; *Vernicia fordii*, water stress.

Introduction

Drought is a major environmental factor that limits productivity, distribution, and survivability of plants (Engelbrecht *et al.* 2007). In recent years, along with global climate change, the frequency, duration and severity of drought has increased in many regions of the world (*e.g.*, Anderegg *et al.* 2012, Flato *et al.* 2013). Plants experiencing drought stress often exhibit serious physiological and biochemical dysfunctions including reduced photosynthesis, decreased transpiration rate (*E*), generation of reactive oxygen species, and damage to various cellular structures (Sperry *et al.* 1998, Lawlor 2002, Ramachandra *et al.* 2004, Xu *et al.* 2008, Niu and Rodriguez 2009, Soares-Cordeiro *et al.* 2009). However, plants adopt different strategies in order to adapt to drought conditions. For example, stomata closure is considered a primary mechanism for regulating water content when

plants are under drought conditions (de Souza *et al.* 2005, Kowitcharoen *et al.* 2015, Zhao *et al.* 2015). Similarly, endogenous abscisic acid and ascorbic acid concentrations in leaves and shoots increase in response to drought stress (Synková and Valcke 2001, Kowitcharoen *et al.* 2015). Many authors reported that drought stress causes a decrease in the chlorophyll (Chl) content, maximal quantum yield of PSII photochemistry (F_v/F_m), and electron transport rate (ETR), but increases dark respiration in wheat (Ahmed *et al.* 2002, Nikolaeva *et al.* 2010, Akhkhia *et al.* 2011, Santos and Silva 2015).

Plant biologists have used various chemicals, such as calcium chloride (Upadhyaya *et al.* 2011, Xu *et al.* 2013), plant growth regulators (Shan *et al.* 2010, Hojati *et al.* 2011), and other substances (Zhao *et al.* 2007, Ishibashi *et al.* 2011) to alleviate the harmful effects of drought stress.

Received 22 April 2016, accepted 14 October 2016, published as online-first 28 November 2016.

[†]Corresponding author; phone: +86-0731-85623416, fax: +86-0731-85623416, e-mail: tanxiaofengcn@126.com

Abbreviations: C_i – intercellular CO₂ concentration; Chl – chlorophyll; DM – dry mass; DAT – day of treatment; *E* – transpiration rate; ETR – electron transport rate; FM – fresh mass; F₀ – minimal fluorescence yield of the dark-adapted state; F_v/F_m – maximal quantum yield of PSII photochemistry; g_s – stomatal conductance; PEPC – phosphoenolpyruvate carboxylase; P_N – net photosynthetic rate; RWC – relative leaf water content; SPAD values – corresponding to content of chlorophyll (*a+b*); SWC – soil water content; Φ_{PSII} – effective quantum yield of PSII.

Acknowledgments: This work was supported by the National Forestry Public Welfare Industry Research Project of China (201204403), a Hunan Provincial Innovation Foundation for Postgraduates (CX2015B286).

Several studies have shown that calcium is involved in the regulation of plant responses under adverse environmental conditions related to drought (Ma *et al.* 2005, Shao *et al.* 2008, Xu *et al.* 2013), heat (Bamberg *et al.* 1998, Wang and Li 2006, Bhattacharjee 2008), salt (Arshi *et al.* 2006, Amor *et al.* 2010), and heavy metals (Antosiewicz and Hennig 2004, Siddiqui *et al.* 2011). Previous studies indicated calcium appears to play an important role in plant defense mechanisms against drought stress. Cousson (2009) revealed Ca^{2+} -signaling was required for the acquisition of drought resistance in *Arabidopsis*. Xu *et al.* (2013) reported calcium-induced drought tolerance in *Zoysia japonica* and increased biomass, Chl content, net photosynthetic rate (P_N), and antioxidant enzyme activities.

Tung tree (*Vernicia fordii* Hemsley) is a member of the Euphorbiaceae family, a native tree species which has been cultivated for more than 1,000 years in China. Along with oil-tea tree (*Camellia oleifera*), walnut (*Juglans regia*), and Chinese tallow tree (*Sapium sebiferum*), it is one of the four major woody oil trees in China (Tan *et al.* 2011). Tung oil, which is extracted from tung seeds, exhibits traits that are highly valued by the industry. Tung oil has fast drying properties, is lightweight, and has excellent

adhesion and glossiness. Additionally, tung oil is resistant to heat, acids, alkalines, and cracking as a result of frost. It is resistant to many chemical treatments, and – unlike other drying oils – it does not darken with age. These properties make tung oil a valuable drying ingredient of paints, varnishes, and other coatings and finishes (Cao and Shockey 2012). Tung tree is a promising biomass species for resolving energy shortage problems (Tan *et al.* 2011). However, various studies indicate water-use efficiency, Chl content and production efficiency decrease in tung tree when subjected to abiotic stresses, especially drought stress (Zhou *et al.* 2012, Li and Zhu 2014). Thus, the growth and yield of tung tree are affected by drought stress. However, only a few studies have investigated the effects of CaCl_2 on the physiology of tung tree under drought.

In this experiment, we studied the effects of exogenous calcium on physiology and growth of one-year-old tung tree seedlings under drought conditions. Our primary objective was to explore the mechanism behind Ca^{2+} -induced alleviation of drought stress on the photosynthetic process in tung tree. For this purpose, we measured photosynthesis as well as the Chl fluorescence parameters and Chl content by SPAD value of tung tree.

Materials and methods

Plant materials and treatments: Tung tree seedlings (*Vernicia fordii* Hemsley) were cultivated in plastic containers (15 cm × 15 cm) containing wet sand under natural conditions at Central South University of Forestry and Technology, Changsha, China. When the seedlings formed a true leaf, they were transplanted to (15 cm × 15 cm) plastic pots filled with a mixture of commercial soil and sand (1:1 ratio) in a nonopaque rain-shelter greenhouse for 1 month before starting the experiment. During the acclimation period, all seedlings were watered 3–4 times per week and fertilized once a week with Hoagland's solution. Once the experiment began, fertilization was stopped.

A total of 72 tung tree seedlings were used in the experiment, which were randomly divided into four groups; each treatment group consisted of three replicates of 18 plants. Aqueous 2, 5, 10 mM CaCl_2 solutions (AR, SCR, Shanghai, China) were sprayed on the leaves of three groups until run-off twice a day for 3 d. The control plants were similarly sprayed with distilled water to run-off. Drought was induced by withholding water for 12 d after CaCl_2 application (DAT). Thus, the treatments were as follows:

Treatment	Specification
Control	No CaCl_2 and drought
2 mM	2 mM CaCl_2 pretreatment and drought
5 mM	5 mM CaCl_2 pretreatment and drought
10 mM	10 mM CaCl_2 pretreatment and drought.

On the 0, 4, 8, 12 DAT, photosynthesis and Chl fluorescence parameters were measured. Immediately, after photosynthesis measurements, the leaves were cut, weighed, wrapped up in tin foil, frozen in liquid nitrogen, and stored at -80°C until analysis. The collected leaf sections were used for measurements of enzyme activities.

Relative leaf water content and soil water content: The relative leaf water content (RWC) was determined at each leaf developmental stage according to the following equation: $\text{RWC} = [(\text{FM} - \text{DM})/(\text{SM} - \text{DM})] \times 100\%$. An analytical balance (0.1 mg precision) (CP522C, OHAUS, Shanghai, China) was used to record the fresh mass (FM), saturated mass (SM), and dry mass (DM) of leaves. The saturated mass was obtained after 24 h of leaf immersion in distilled water in the dark. The DM was obtained after drying leaves in an oven (DHG-9202, SANFA, Shanghai, China) for 48 h at 60°C . Soil water content (SWC) was determined after the treatment period according to the following equation: $\text{SWC} = [(\text{soil wet mass}) - (\text{soil dry mass})]/(\text{soil wet mass}) \times 100\%$. After the treatment period was complete, the soil wet mass was weighed immediately. The soil DM was obtained after drying soil in an oven (DHG-9202, SANFA, Shanghai, China) for 48 h at 80°C .

Determination of Ca^{2+} content and relative Chl content: The leaves were dried in an oven at 105°C for 30 min and ground after drying at 65°C to a constant mass. The content of Ca^{2+} in leaves of tung tree was measured according to the method described in Chinese National

Standard LY/T 1270–1999. Ca^{2+} was measured by atomic absorption spectrometry (TAS-990, PGENERAL, Beijing, China). Relative Chl content was measured using a SPAD meter (SPAD-502, Minolta, Tokyo, Japan).

Enzyme extraction and assays: Fresh leaves (0.5 g) were ground with 3 mL of ice-cold 100 mM Tris-HCl buffer (pH 8.2) (T8230, Solarbio, Beijing, China) containing 5% glycerin, 1 mM EDTA (Amresco0105, Solarbio, Beijing, China), 7 mM mercaptoethanol, and 1% PVP (K-30, Solarbio, Beijing, China). The homogenates were centrifuged at 4°C for 20 min at $15,000 \times g$ and the supernatant was stored at 4°C for enzyme activity assay. The carbon assimilatory enzymes Rubisco (EC 4.1.1.39) and phosphoenolpyruvate carboxylase (PEPC, EC 4.1.1.31) were assayed according to Siegel (1975) and Shi *et al.* (1979), respectively. The absorbance was measured at 340 nm by spectrophotometer (UV 9100 D, LabTech, Beijing, China) and activity was expressed as $\mu\text{mol}(\text{CO}_2) \text{ min}^{-1} \text{ g}^{-1}(\text{FM})$.

Leaf gas exchange was measured on the uppermost fully expanded leaves using a LI-6400XT portable photosynthesis system (LI-COR, Lincoln, NE, USA) during midday (9:00–11:00 h) at each measurement period (one

leaf per plant; six plants per replicate). Light was provided by LEDs emitting in the blue-red light spectrum with a constant saturating light intensity of $1,500 \mu\text{mol}(\text{photon}) \text{ m}^{-2} \text{ s}^{-1}$ and an ambient CO_2 concentration of $400 \mu\text{mol} \text{ mol}^{-1}$. The temperature of the leaf cuvette was 30°C and the relative humidity was 70%.

Measurement of Chl fluorescence: Fluorescence parameters were measured with LI-6400-40LCF (LI-COR, Lincoln, NE, USA). Three leaves of each treatment were dark-adapted for 30 min prior to the fluorescence measurements. The minimal Chl fluorescence (F_0) level was measured after applying a far-red pulse for 6 s and the maximal fluorescence (F_m) was registered after applying a 0.8 s saturating flash. Maximal photochemical efficiency of PSII (F_v/F_m) was expressed as: $F_v/F_m = (F_m - F_0)/F_m$ (Genty *et al.* 1989).

Statistical analysis: All data were obtained from three replicates and the results presented were the mean values. All data were subjected to one way analysis of variance (ANOVA) with SPSS 17.0 software. When the main effect was a significant, Duncan's multiple range test was performed at the 0.05 level of significance.

Results

Leaf relative water content and soil water content: The RWC values showed significant reduction in control groups at 8 and 12 DAT (Fig. 1A). Compared with the control, at 12 DAT, leaf RWC of plants pretreated with 5 mM CaCl_2 significantly increased by 11.4%; however, no significant differences were found between concentrations of 2, 5, and 10 mM CaCl_2 during the whole experiment (Fig. 1A). The SWC decreased significantly, as expected, as drought was prolonged, and there was no difference

observed between the CaCl_2 pretreatment group and the control (Fig. 1B).

Ca^{2+} content and relative Chl content: Application of CaCl_2 had no significant advantage for Ca^{2+} accumulation compared with the control at the beginning of drought stress, while the CaCl_2 pretreatment resulted in an increase in Ca^{2+} content at 4 DAT (Fig. 2A). At 8 and 12 DAT, the plants pretreated with CaCl_2 demonstrated similar Ca^{2+}

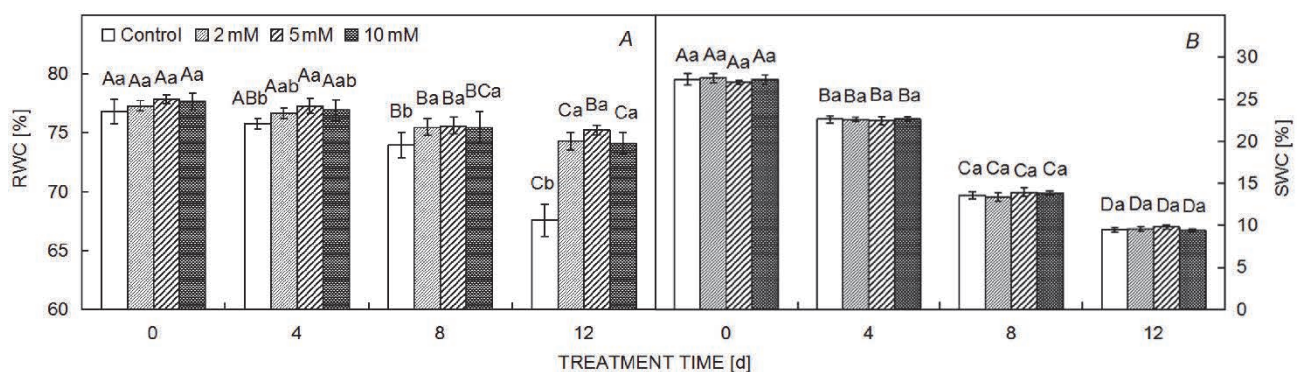


Fig. 1. Effects of CaCl_2 on RWC (A) in tung tree seedlings and SWC (B) under drought stress. (1) No CaCl_2 and drought; control, (2) 2 mM CaCl_2 pretreatment and drought; 2 mM, (3) 5 mM CaCl_2 pretreatment and drought; 5 mM, and (4) 10 mM CaCl_2 pretreatment and drought; 10 mM. The data in the figure represent mean $\pm$ SE ($n = 3$). Different capital letters indicate statistical differences between drought treatment stages, and lowercase letters indicate significant difference between Ca treatments at the same drought treatment stage, at $P \leq 0.05$ according to Duncan's multiple range test (DMRT).

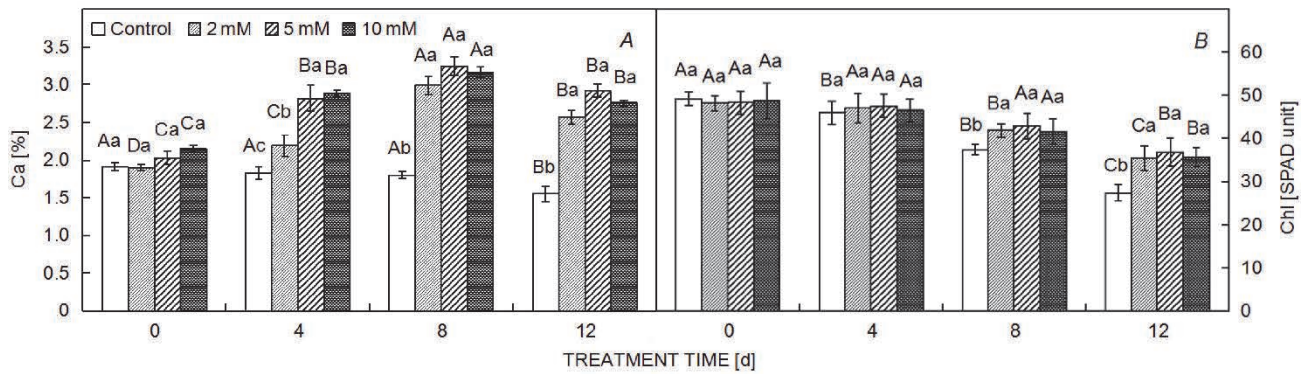


Fig. 2. Effects of CaCl_2 on Ca content, Chl SPAD value in leaves of tung tree seedlings under drought stress. (1) No CaCl_2 and drought; control, (2) 2 mM CaCl_2 pretreatment and drought; 2 mM, (3) 5 mM CaCl_2 pretreatment and drought; 5 mM, and (4) 10 mM CaCl_2 pretreatment and drought; 10 mM. The data in the figure represent mean $\pm$ SE ($n = 3$). Different capital letters indicate statistical differences between drought treatment stages, and lowercase letters indicate significant difference between Ca treatments at the same drought treatment stage, at $P \leq 0.05$ according to Duncan's multiple range test (DMRT).

contents, which were 80 to 88% higher, respectively, than that of the control. The SPAD value in all pretreatments decreased as the drought stress persisted (Fig. 2B). However, the CaCl_2 pretreatment alleviated the loss of Chl induced by water stress. Compared with the stressed control, SPAD values significantly increased in the CaCl_2 -pretreated plants at 8 and 12 DAT. Meanwhile, no significant differences in SPAD values were observed in CaCl_2 treatments and stressed control at 4 DAT.

Rubisco and PEPC: The drought stress decreased the activity of both Rubisco and PEPC enzymes and this effect became more pronounced with time (Fig. 3). The plants in all groups pretreated with CaCl_2 overcame the drought-induced reduction in activities of the CO_2 -assimilatory enzymes, which was especially evident at 8 and 12 DAT. Compared with the control, Rubisco and PEPC activities

of the tung tree leaves increased in the CaCl_2 -pretreated plants at 8 and 12 DAT.

Leaf gas exchange: The P_N , g_s , and E of all treatments and the control decreased as the drought stress continued. However, the C_i of the control first decreased and then increased, while the CaCl_2 -pretreated plants exhibited no obvious changes except for 8 DAT (Fig. 4). At 0 DAT, there was no significant difference between the CaCl_2 pretreatment and control in P_N and E , while the plants pretreated with 5 mM CaCl_2 showed a dramatic increase in g_s , compared with the control and other treatments. Meanwhile, g_s of the CaCl_2 -pretreated plants was significantly higher than that of the control at 4 DAT and g_s of plants pretreated with 5 mM CaCl_2 was higher than the control at 8 and 12 DAT. Plants pretreated with 2 mM CaCl_2 did not show significant differences in g_s compared with the control.

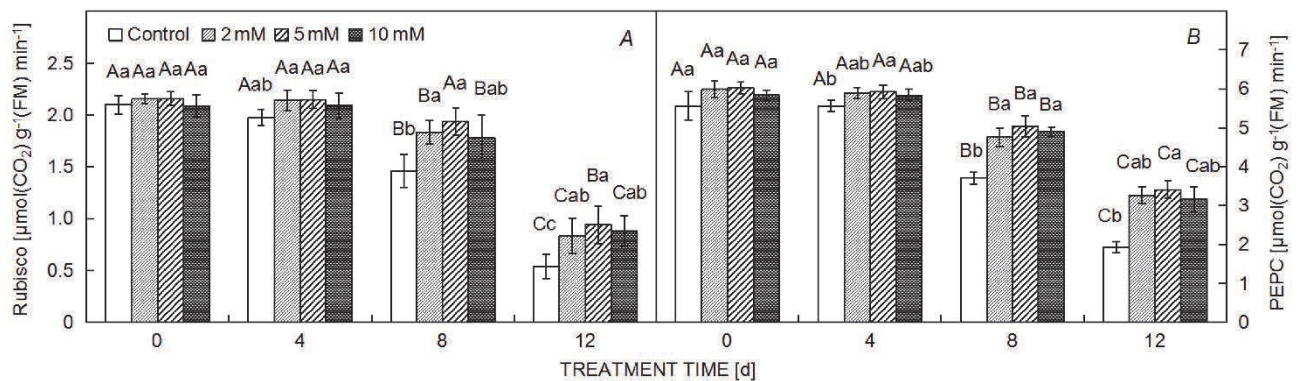


Fig. 3. Effects of CaCl_2 on enzymatic activity of Rubisco and phosphoenolpyruvate carboxylase (PEPC) in tung tree seedlings under drought stress. (1) No CaCl_2 and drought; control, (2) 2 mM CaCl_2 pretreatment and drought; 2 mM, (3) 5 mM CaCl_2 pretreatment and drought; 5 mM, and (4) 10 mM CaCl_2 pretreatment and drought; 10 mM. The data in the figure represent mean $\pm$ SE ($n = 3$). Different capital letters indicate statistical differences between drought treatment stages, and lowercase letters indicate significant difference between Ca treatments at the same drought treatment stage, at $P \leq 0.05$ according to Duncan's multiple range test (DMRT).

Chl fluorescence: Drought stress increased the F_0 in tung tree leaves 8 and 12 DAT after withholding irrigation (Fig. 5A). However, F_v/F_m , Φ_{PSII} , and ETR of plants without CaCl_2 pretreatment decreased gradually with time under drought stress (Fig. 5B–D). No difference was detected between CaCl_2 pretreatment and control for F_v/F_m

and Φ_{PSII} at 0 and 4 DAT (Fig. 5B,C). In addition, 5 mM CaCl_2 pretreatment increased ETR compared with the control group. Drought stress markedly reduced both F_v/F_m and Φ_{PSII} at 12 DAT, and the values of F_v/F_m and Φ_{PSII} in plants with CaCl_2 pretreatment, except for those at 2 mM CaCl_2 , were higher than that of the control.

Discussion

RWC is an appropriate estimate of plant water status in terms of cellular hydration under drought conditions. RWC started to decrease at 4 DAT, but there was no statistical difference until 8 DAT, while SWC decreased by 49.9 and 64.9% on 4 and 8 DAT, respectively, compared to that on 0 DAT. On 8 DAT, leaf RWC in the plants with CaCl_2 pretreatment was 2.2% higher than that of the control, indicating ameliorative effect of the CaCl_2 pretreatment. With the proceeding stress, SWC decreased by 9.6%, RWC in the plants pretreated with CaCl_2 was 12.0% higher than the control at 12 DAT, but was 2.9 % lower than that on 0 DAT. The results indicated that RWC of tung tree seedlings improved by the CaCl_2 pretreatment under drought stress.

Calcium is involved in the regulatory mechanisms that help plants adjust to adverse environmental conditions and play an important role in maintaining the stability of cell membrane phospholipids and proteins (Ma *et al.* 2005, Kim *et al.* 2009, Upadhyaya *et al.* 2011). In this study, Chl SPAD values at 8 and 12 DAT were reduced by drought because of the destruction of pigments and the instability of the pigment-protein complexes (Levitt 1980, Xu *et al.* 2013); however, exogenous CaCl_2 application alleviated the drought effect (Fig. 2B). This result is similar to observations found in zoysiagrass (*Zoysia japonica*), where Chl content was enhanced by CaCl_2 pretreatment under drought stress (Xu *et al.* 2013). At 4 DAT, the P_N and g_s with CaCl_2 pretreatment were higher than the control, while the carboxylation of both Rubisco and PEPC did not increase compared with the control until 8 and 12 DAT. The main reason was that drought stress led to a slight stomatal closure at 4 DAT, which decreased the P_N . Along with the increase of drought stress, both Rubisco and PEPC increased in the plants pretreated with CaCl_2 at 8 and 12 DAT compared with the control. The results indicated that Ca^{2+} could alleviate stress-induced damages and increase photosynthetic performance in tung tree. One reason might be that the Ca^{2+} treatment prevented the dehydration damage of cellular structure by maintaining the osmotic strength of the cytoplasm in plants (Gzik 1996, Arshi *et al.* 2006, Yang *et al.* 2016).

Many studies have indicated that CaCl_2 could act as a physiological treatment to increase plant drought tolerance (Tan *et al.* 2011, Xu *et al.* 2013, Yang *et al.* 2016). Barbosa *et al.* (2015) reported that the carboxylation of Rubisco and PEPC were the main limiting factors for photosynthesis under drought stress. Synková *et al.* (2006) reported that reduction of P_N was caused by the decrease of Rubisco

activity and stomata closure. The results of our study showed that the carboxylation activity of both Rubisco and PEPC was significantly reduced at 8 to 12 DAT (Fig. 3). Drought stress significantly reduced P_N and E in the control group, whereas exogenous CaCl_2 application diminished this decrease (Fig. 4A,D). The change of g_s in the plants pretreated with CaCl_2 exhibited a similar trend, but g_s significantly decreased after 4 DAT. However, exogenous CaCl_2 application reduced the decrease (Fig. 4B).

Decreasing C_i and the inhibition of Rubisco enzyme activity and ATP synthesis lead to a decrease of P_N (Dulai *et al.* 2006). The present investigation indicated that the CaCl_2 pretreatment could improve the P_N of tung tree under drought stress conditions. Many studies have indicated that CaCl_2 could increase plant tolerance, especially to drought stress (Ma *et al.* 2005, Xu *et al.* 2013). Ca^{2+} may regulate stomata movement, decrease respiration intensity, and activate the NAD kinase, prompting NADP production processes (Jones *et al.* 1967). Under a water deficit, there are stomatal and nonstomatal limitations of photosynthesis; when g_s and C_i decrease at the same time, the P_N decline is mainly caused by stomatal limitation. If g_s decreases while C_i increases, the photosynthesis is limited mainly by nonstomatal factors (Farquhar and Sharkey 1982). The simultaneous decline of P_N , g_s , and C_i in the control group at 4 DAT clearly indicated that the stomata closure was the main factor responsible for reduction in the P_N . Later on, after 12 DAT under drought, the rise in C_i let us hypothesize that the drop in P_N was mainly due to nonstomatal factors, such as a reduction of CO_2 -assimilatory enzyme activity. We found that at the beginning of drought stress, 5 mM CaCl_2 -treated leaves had higher g_s than those of other treatments, but P_N and E did not increase significantly. The increased g_s in this study by the CaCl_2 pretreatment may be due to opening of stomatal aperture as reported by Allen *et al.* (2001).

Chl fluorescence is an indicator of PSII functioning and electron transfer from PSII to PSI and has been used frequently to estimate damage to the photosynthetic system (Niu *et al.* 2008). An increase in F_0 was observed on 8 DAT only for the control, and the F_0 with CaCl_2 pretreatment were lower than the control after 12 DAT. Our results showed that 5 mM CaCl_2 treatment significantly increased F_v/F_m and Φ_{PSII} in tung tree seedlings after 8 DAT of drought stress (Fig. 5B,C). In this study, we found that CaCl_2 pretreatment significantly increased ETR from 8 to 12 DAT. This finding was consistent with previous studies, which demonstrated that Ca^{2+} could

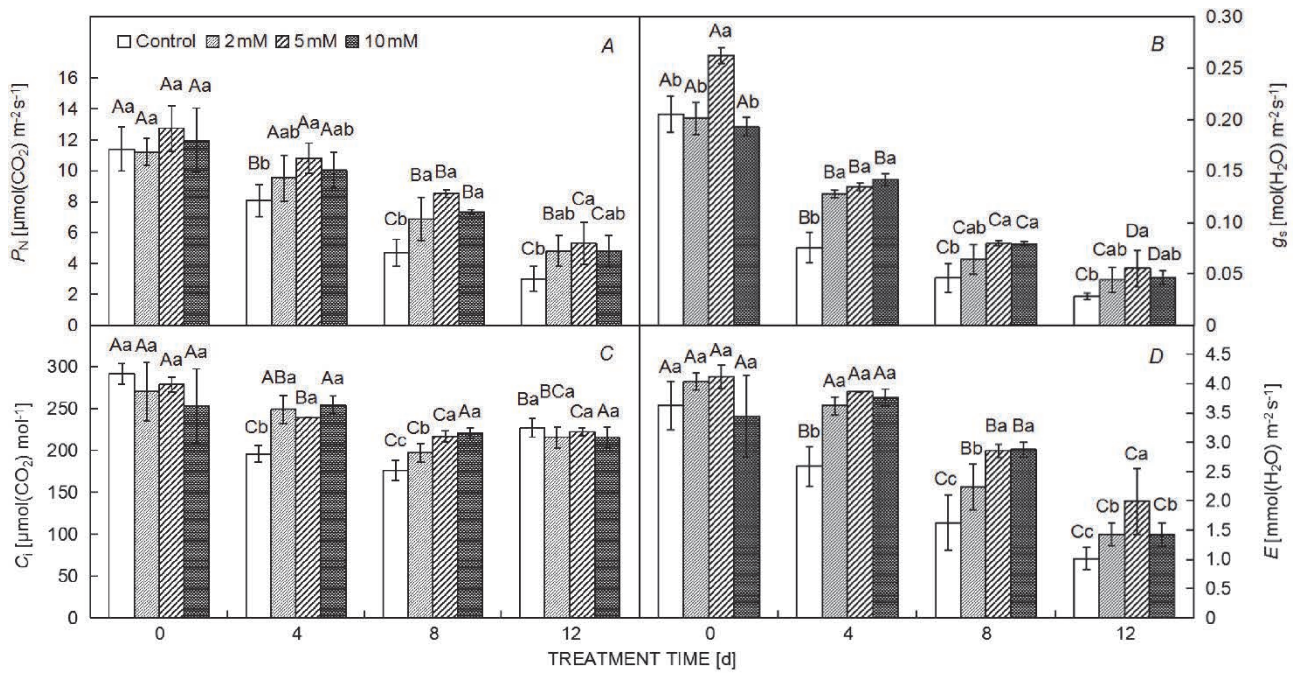


Fig. 4. Effects of CaCl_2 on net photosynthetic rate (P_N), stomatal conductance (g_s), intercellular CO_2 concentration (C_i), and transpiration rate (E) in tung tree seedlings under drought stress. (1) No CaCl_2 and drought; control, (2) 2 mM CaCl_2 pretreatment and drought; 2 mM, (3) 5 mM CaCl_2 pretreatment and drought; 5 mM, and (4) 10 mM CaCl_2 pretreatment and drought; 10 mM. The data in the figure represent mean $\pm \text{SE}$ ($n = 3$). Different capital letters indicate statistical differences between drought treatment stages, and lowercase letters indicate significant difference between Ca treatments at the same drought treatment stage, at $P \leq 0.05$ according to Duncan's multiple range test (DMRT).

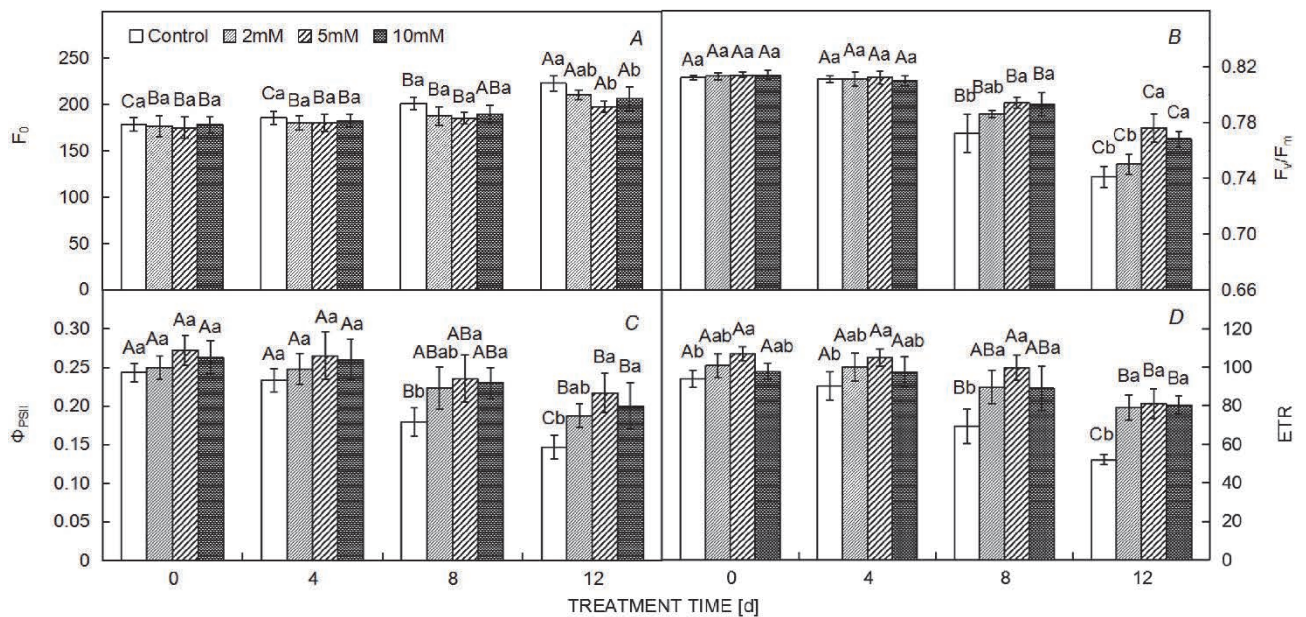


Fig. 5. Effects of CaCl_2 on minimal fluorescence yield of the dark-adapted state (F_0), maximal quantum yield of PSII photochemistry (F_v/F_m), effective quantum yield of PSII (Φ_{PSII}) and electron transport rate (ETR) in tung tree seedlings under drought stress. (1) No CaCl_2 and drought; control, (2) 2 mM CaCl_2 pretreatment and drought; 2 mM, (3) 5 mM CaCl_2 pretreatment and drought; 5 mM, and (4) 10 mM CaCl_2 pretreatment and drought; 10 mM. The data in the figure represent mean $\pm \text{SE}$ ($n = 3$). Different capital letters indicate statistical differences between drought treatment stages, and lowercase letters indicate significant difference between Ca treatments at the same drought treatment stage, at $P \leq 0.05$ according to Duncan's multiple range test (DMRT).

partially alleviate PSII reaction center closure and improve utilization of light energy (Wei *et al.* 2015). Previous studies showed that different CaCl_2 concentrations significantly influenced plant tolerance of drought stress, which indicated F_v/F_m was higher in *Zoysia japonica* pretreated with 10 mM CaCl_2 than in other treatments (Xu *et al.* 2013). Amor *et al.* (2010) reported that 3.5 mM Ca^{2+} was more effective than 20 mM in lowering NaCl stress for *Cakile maritima*. These studies indicate that different CaCl_2 concentrations result in different levels of plant tolerance towards abiotic stresses. Compared to 2 and 10 mM, we found that 5 mM Ca^{2+} relieved the most effectively the injury of drought stress.

References

- Ahmed S., Nawata E., Hosokawa M. *et al.*: Alterations in photosynthesis and some antioxidant enzymatic activities of mungbean subjected to waterlogging. – *Plant Sci.* **163**: 117-123, 2002.
- Akhkha A., Boutraa T., Alhejely A.: The rates of photosynthesis, chlorophyll content, dark respiration, proline and abscisic acid (ABA) in wheat (*Triticum durum*) under water deficit conditions. – *Int. J. Agric. Biol.* **13**: 215-221, 2011.
- Allen G.J., Chu S.P., Harrington C.L. *et al.*: A defined range of guard cell calcium oscillation parameters encodes stomatal movements. – *Nature* **411**: 1053-1057, 2001.
- Amor N.B., Megdiche W., Jiménez A. *et al.*: The effect of calcium on the antioxidant systems in the halophyte *Cakile maritima* under salt stress. – *Acta Physiol. Plant.* **32**: 453-461, 2010.
- Anderegg W.R.L., Kane J.M., Anderegg L.D.L.: Consequences of widespread tree mortality triggered by drought and temperature stress. – *Nat. Clim. Change* **3**: 30-36, 2012.
- Antosiewicz D.M., Hennig J.: Overexpression of *LCT1* in tobacco enhances the protective action of calcium against cadmium toxicity. – *Environ. Pollut.* **129**: 237-245, 2004.
- Arshi A., Abidin M.Z., Iqbal M.: Effect of CaCl_2 on growth performance, photosynthetic efficiency and nitrogen as assimilation of *Cichorium intybus* L. grown under NaCl stress. – *Acta Physiol. Plant.* **28**: 137-147, 2006.
- Bamberg J.B., Palta J.P., Peterson L.A. *et al.*: Fine screening potato (*Solanum*) species germplasm for tuber calcium. – *Am. J. Potato Res.* **75**: 181-186, 1998.
- Barbosa A.M., Guidorizi K.A., Catuchi T.A. *et al.*: Biomass and bioenergy partitioning of sugarcane plants under water deficit. – *Acta Physiol. Plant.* **37**: 142-149, 2015.
- Bhattacharjee S.: Calcium-dependent signaling pathway in the heat-induced oxidative injury in *Amaranthus lividus*. – *Biol. Plantarum* **52**: 137-140, 2008.
- Cao H.P., Shockey J.M.: Comparison of TaqMan and SYBR green qPCR methods for quantitative gene expression in tung tree tissues. – *J. Agr. Food Chem.* **60**: 12296-12303, 2012.
- Cousson A.: Involvement of phospholipase C-independent calcium-mediated abscisic acid signaling during *Arabidopsis* response to drought. – *Biol. Plantarum* **53**: 53-62, 2009.
- de Souza C.R., Maroco J.P., dos Santos T.P. *et al.*: Control of stomatal aperture and carbon uptake by deficit irrigation in two grapevine cultivars. – *Agr. Ecosyst. Environ.* **106**: 261-274, 2005.
- dos Santos C.M., Silva M.D.A.: Physiological and biochemical responses of sugarcane to oxidative stress induced by water deficit and paraquat. – *Acta Physiol. Plant.* **37**: 172, 2015.
- Dulai S., Molnár I., Prónay J. *et al.*: Effects of drought on photosynthetic parameters and heat stability of PSII in wheat and in *Aegilops* species originating from dry habitats. – *Acta Biol. Szeged.* **50**: 11-17, 2006.
- Engelbrecht B.M.J., Comita L.S., Condit R. *et al.*: Drought sensitivity shapes species distribution patterns in tropical forests. – *Nature* **447**: 80-82, 2007.
- Farquhar G.D., Sharkey T.D.: Stomatal conductance and photosynthesis. – *Annu. Rev. Plant Physiol.* **33**: 317-345, 1982.
- Flato G., Marotzke J., Abiodun B. *et al.*: Evaluation of climate models. – In: *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Pp. 741-866. Cambridge Univ. Press, Cambridge and New York 2013.
- Genty B., Briantais J.M., Baker N.R.: The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. – *Biochim. Biophys. Acta.* **990**: 87-92, 1989.
- Gzik A.: Accumulation of proline and pattern of amino acids in sugarbeet plants in response to osmotic, water and salt stress. – *Environ. Exp. Bot.* **36**: 29-38, 1996.
- Hojati M., Modarres-Sanavy S.A.M., Ghanati F. *et al.*: Hexaconazole induces antioxidant protection and apigenin-7-glucoside accumulation in *Matricaria chamomilla* plants subjected to drought stress. – *J. Plant Physiol.* **168**: 782-791, 2011.
- Ishibashi Y., Yamaguchi H., Yuasa T. *et al.*: Hydrogen peroxide spraying alleviates drought stress in soybean plants. – *J. Plant Physiol.* **168**: 1562-1567, 2011.
- Jones R.G., Lunt O.R.: The function of calcium in plants. – *Bot. Rev.* **33**: 407-426, 1967.
- Kim M.C., Chung W.S., Yun D.J. *et al.*: Calcium and calmodulin-mediated regulation of gene expression in plants. – *Mol. Plant.* **2**: 13-21, 2009.
- Kowitcharoen L., Wongs-Aree C., Setha S. *et al.*: Changes in abscisic acid and antioxidant activity in sugar apples under drought conditions. – *Sci. Hortic.-Amsterdam* **193**: 1-6, 2015.
- Lawlor D.W.: Limitations to photosynthesis in water stressed leaves: stomata vs. metabolism and the role of ATP. – *Ann. Bot.-London* **89**: 871-885, 2002.
- Levitt J.: Responses of Plants to Environmental Stresses. Pp. 3642-3645. Academic Press, New York 1980.

- Li Q.M., Zhu Z.P.: [Ecological factors affecting the growth and fruiting of *Vernicia fordii* in Shangnan county.] – Shanxi Forest Sci. Tech. **3**: 82-84, 2014. [In Chinese].
- Ma R., Zhang M., Li B. *et al.*: The effects of exogenous Ca^{2+} on endogenous polyamine levels and drought resistant traits of spring wheat grown under arid conditions. – J. Arid. Environ. **63**: 177-190, 2005.
- Nikolaeva M.K., Maevskaya S.N., Shugaev A.G. *et al.*: Effect of drought on chlorophyll content and antioxidant enzyme activities in leaves of three wheat cultivars varying in productivity. – Russ. J. Plant Physiol. **57**: 87-95, 2010.
- Niu G.H., Rodriguez D.S.: Growth and physiological responses of four rose rootstocks to drought stress. – J. Am. Soc. Hortic. Sci. **134**: 202-209, 2009.
- Niu G.H., Rodriguez D.S., Mackay W.: Growth and physiological responses to drought stress in four oleander clones. – J. Am. Soc. Hortic. Sci. **133**: 188-196, 2008.
- Ramachandra Reddy A., Chaitanya K., Jutur P. *et al.*: Differential antioxidative responses to water stress among five mulberry (*Morus alba* L.) cultivars. – Environ. Exp. Bot. **52**: 33-42, 2004.
- Shan C.J., Liang Z.S.: Jasmonic acid regulates ascorbate and glutathione metabolism in *Agropyron cristatum* leaves under water stress. – Plant Sci. **178**: 130-139, 2010.
- Shao H.B., Song W.Y., Chu L.Y.: Advances of calcium signals involved in plant anti-drought. – C.R. Biol. **331**: 587-596, 2008.
- Shi J.N., Wu M.X., Cha J.J.: [Studies on plant phosphoenolpyruvate carboxylase I. Separation and properties of PEP carboxylase isoenzymes.] – J. Plant Physiol. **5**: 225-236, 1979. [In Chinese]
- Siddiqui M.H., Al-Whaibi M.H., Basalah M.O.: Interactive effect of calcium and gibberellin on nickel tolerance in relation to antioxidant systems in *Triticum aestivum* L. – Protoplasma **248**: 503-511, 2011.
- Siegel M.I., Lane M.D.: Ribulose-diphosphate carboxylase from spinach leaves. – Methods Enzymol. **42**: 472-480, 1975.
- Soares-Cordeiro A.S., Carmo-Silva A.E., Bernardes da Silva A. *et al.*: Effects of rapidly imposed water deficit on photosynthetic parameters of three C_4 grasses. – Photosynthetica **47**: 304-308, 2009.
- Sperry J.S., Adler F.R., Campbell G.S. *et al.*: Limitation of plant water use by rhizosphere and xylem conductance: results from a model. – Plant Cell Environ. **21**: 347-359, 1998.
- Synková H., Valcke R.: Response to mild water stress in transgenic *Pssu-ipt* tobacco. – Physiol. Plant. **112**: 513-523, 2001.
- Synková H., Semorádová Š., Schnablová R. *et al.*: Effects of biotic stress caused by *Potato virus Y* on photosynthesis in *ipt* transgenic and control *Nicotiana tabacum* L. – Plant Sci. **171**: 607-616, 2006.
- Tan X.F., Jiang G.X., Tan F.Y. *et al.*: [Research report on industrialization development strategy *Vernicia fordii* of in China.] – Nonwood Forest Res. **29**: 1-5, 2011. [In Chinese].
- Upadhyaya H., Panda S.K., Dutta B.K.: CaCl_2 improves post drought recovery potential in *Camellia sinensis* (L.) O. Kuntze. – Plant Cell Rep. **30**: 495-503, 2011.
- Wang L.J., Li S.H.: Salicylic acid-induced heat or cold tolerance in relation to Ca^{2+} homeostasis and antioxidant systems in young grape plants. – Plant Sci. **170**: 685-694, 2006.
- Wei R., Liu Y., Sui Y. *et al.*: Effects of Ca^{2+} and polyethylene glycol on the chlorophyll fluorescence parameters of transgenic *Oryza sativa* rice (*Oryza sativa* L.). – Photosynthetica **53**: 336-341, 2015.
- Xu C.B., Li X.M., Zhang L.H.: The effect of calcium chloride on growth, photosynthesis, and antioxidant responses of *Zoysia japonica* under drought conditions. – PLoS ONE **8**: 0068214, 2013.
- Xu X., Peng G., Wu C. *et al.*: Drought inhibits photosynthetic capacity more in females than in males of *Populus cathayana*. – Tree Physiol. **28**: 1751-1759, 2008.
- Yang B.Z., Liu Z.B., Zhou S.D. *et al.*: Exogenous Ca^{2+} alleviates waterlogging-caused damages to pepper. – Photosynthetica **54**: 620-629, 2016.
- Zhao H.J., Tan J.F., Qi C.M.: Photosynthesis of *Rehmannia glutinosa* subjected to drought stress is enhanced by choline chloride through alleviating lipid peroxidation and increasing proline accumulation. – Plant Growth Regul. **51**: 255-262, 2007.
- Zhao W.S., Sun Y.L., Kjellgren R. *et al.*: Response of stomatal density and bound gas exchange in leaves of maize to soil water deficit. – Acta Physiol. Plant. **37**: 1704-1713, 2015.
- Zhou Y.C., Wang R., Liu Y. *et al.*: [Resistance and stable carbon isotopic composition of two species of biomass plants under osmotic stress.] – Earth Environ. **40**: 485-490, 2012. [In Chinese].