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Sensing and classification of rice (*Oryza sativa* L.) drought stress levels based on chlorophyll fluorescence

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Abstract

Sensing and classification of drought stress levels are very important to agricultural production. In this work, rice drought stress levels were classified based on the commonly used chlorophyll *a* fluorescence (ChlF) parameter (F_v/F_m), feature data (induction features), and the whole OJIP induction (induction curve) by using a Support Vector Machine (SVM). The classification accuracies were compared with those obtained by the K-Nearest Neighbors (KNN) and the Ensemble model (Ensemble) correspondingly. The results show that the SVM can be used to classify drought stress levels of rice more accurately compared to the KNN and the Ensemble and the classification accuracy (86.7%) for the induction curve as input is higher than the accuracy (43.9%) with F_v/F_m as input and the accuracy (72.7%) with induction features as input. The results imply that the induction curve carries important information on plant physiology. This work provides a method of determining rice drought stress levels based on ChlF.

Keywords: chlorophyll *a* fluorescence; drought stress; Ensemble model; K-Nearest Neighbors; Support vector machine.

Introduction

With global climate and environmental changes, the frequency and duration of irregular precipitation and continuous drought stress caused by extreme weather are increasing, and the degree of crop damage and loss is also rising (Webber *et al.* 2018, Xu *et al.* 2020). In addition, pollution further exacerbates the water resource crisis.

As a major crop, rice demands a high level of soil moisture (Carrijo *et al.* 2018). Water is tightly connected with crop photosynthetic capacity, which can seriously impact the yield and quality of rice (Rosegrant and Cline 2003, Xu *et al.* 2020). With the shortage of water, drought stress has become an important factor in reducing rice yield (Iturbe-Ormaetxe *et al.* 1998, Yang *et al.* 2019, Melandri *et al.* 2020). Thus, to improve rice production and reduce

Highlights

- An SVM-based method was developed to classify rice drought stress levels
- All the data on the chlorophyll fluorescence induction OJIP were used as inputs
- Results were more accurate than analysis with only using some feature parameters

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Abbreviations: ChlF – chlorophyll *a* fluorescence; D0 – no drought stress treatment; D1 – drought stress treatment for 1 h; D4 – drought stress treatment for 4 h; KNN – K-Nearest Neighbors; OJIP – chlorophyll *a* fluorescence induction kinetics curve; SVM – Support Vector Machine.

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irrigation costs, it is of great significance to study rice drought stress measures (Sayed 2003).

Photosynthesis is an important physiological process, which is closely related to plant water status, and it is very sensitive to water shortage. Photosynthetic rate decreases under drought stress (Miyashita *et al.* 2005). When leaf water content drops to a certain extent, plant photosynthesis will be inhibited or even completely stopped. Drought events are usually characterized by drought indices (Marcos-Garcia *et al.* 2017, Mukherjee *et al.* 2018). In agriculture, drought monitoring methods can be divided into three categories: meteorological monitoring methods (Jiao *et al.* 2019), remote sensing methods (Kumar *et al.* 2014, West *et al.* 2019), and integrated remote sensing monitoring methods (Haboudane *et al.* 2004, Sánchez *et al.* 2018). However, these methods do not directly relate to photosynthetic efficiency. There is still a lack of universal, noninvasive, convenient, and affordable plant-physiology-based measures of drought stress that can reflect the influence of drought on photosynthesis.

ChlF emission from photosystem II (PSII) couples with light energy absorption, transmission, dissipation, and distribution, and it plays an important role in determining photosynthesis (Lazár 2006, Maghsoudi *et al.* 2015, Urban *et al.* 2017). It can sense the effects of the external environment on the photosynthesis and physiology of plants, even if the changes in the external environment are very subtle. It is widely used in photosynthesis analysis, plant stress physiology research, oceanography, and remote sensing (Feng *et al.* 2018, Xia *et al.* 2018, Fu *et al.* 2019, Wang *et al.* 2019), and even genetic breeding under drought environments (Woo *et al.* 2008, Faraloni *et al.* 2011, Guo and Tan 2015, Zendonadi *et al.* 2021). ChlF measurement does not damage plant cells or cause harm to organisms. Thus, it may be used to measure the effect of drought on rice (Maghsoudi *et al.* 2015).

In the literature, researchers studied the influence of environmental factors on ChlF to indirectly study the influence of the environment on plant photosynthesis. The minimum dark ChlF (F_o), the maximum dark ChlF (F_m), and the parameters calculated according to some feature data on the ChlF induction are often used for analysis (Banks 2018, Meravi and Prajapati 2020). At the same time, most of the data on the measured ChlF induction curve is not used for the analysis of plants; however, it may contain useful information and enhance the signal-to-noise ratio.

Support Vector Machine (SVM) is a very effective classification tool and has been widely used in agricultural plant recognition. SVM algorithm is a supervised learning model defined by separating hyperplanes based on the statistical learning theory. It is a method in the field of artificial intelligence and data mining. The generalization performance of SVM focuses on the principle of structural risk minimization (SRM), rather than minimizing the error in the training data, which thus avoids common overfitting problems in most Artificial Neural Network (ANN) models. Therefore, in an SVM model, even if the dimensionality of the training samples is very high, SVM can obtain high generalization ability. A large number of

applications also show that SVM has good classification capabilities in processing a small number of samples, nonlinear, and high-dimensional pattern recognition.

SVM was widely applied in agriculture, for example, material identification (Karimi *et al.* 2006, Guerrero *et al.* 2012, Kour and Arora 2019, Qiao *et al.* 2019), adulteration percentage classification (Timsorn *et al.* 2017), plant substance content prediction (Ignat *et al.* 2013, Wang *et al.* 2018), and disease classification (Alam and Kwon 2017). Although SVM has been widely used in agricultural plants, it has not been used in plant drought stress recognition based on ChlF.

Drought stress is one of the main environmental factors that restrict rice growth. The damage of drought stress to rice growth is multifaceted and the response mechanism of rice to drought stress is also extremely complex, which can lead to rice yield reduction and even death. The research on rice drought stress response has reached the molecular level, involving cloning, positioning, and functional analysis of drought resistance genes. However, there is a lack of a universal, noninvasive, convenient, and economic method for determining drought stress levels based on plant physiology to reflect the impact of drought stress on photosynthesis. Therefore, in this work, SVM was used to classify rice drought stress levels with the entire OJIP induction curve or computed induction feature(s) as input variables. In addition, the K-Nearest Neighbors (KNN) and the Ensemble methods were used for comparison.

Materials and methods

Experimental samples: Rice plants (*Oryza sativa* L.) were uprooted from a production field at 6:30 h in July 2019 after growing for about 40 d. To reduce initial differences in moisture contents, the roots of rice were immersed in water for two hours. Then the roots were put into a 20% polyethylene glycol (PEG) solution for different durations (D0 – no drought stress treatment; D1 – drought stress treatment for 1 h; D4 – drought stress treatment for 4 h) to achieve different levels of drought stress. PEG-induced drought is commonly used in plant drought stress research (Wang *et al.* 2005, Cai *et al.* 2015, Faseela *et al.* 2020). We used 883 rice plants.

Fluorescence measurement: ChlF of the treated plant samples was measured with a *FluorPen* hand-held fluorometer (Model FF 110, Photon Systems Instruments, Czech Republic) set to its OJIP protocol.

Data analysis: The measured ChlF induction responses were used as inputs and the drought level (D0, D1, and D4) as the classifier or output. The induction responses were used in three ways: (1) a ChlF parameter: maximum photochemical quantum yield of PSII in the dark (F_v/F_m); (2) induction features: computed features of ChlF induction (F_o , F_j , F_i , F_m , F_v , V_j , V_i , F_m/F_o , F_v/F_o , F_v/F_m , M_0 , Area, Fix Area, S_m , S_s , ϕP_0 , ψ_0 , ϕE_0 , ϕD_0 , ϕPav , PI_{ABS} , ABS/RC , TR_o/RC , ET_o/RC , and DI_o/RC) (see Appendix for a more detailed explanation about ChlF parameters);

(3) induction curve: all the data of the ChlF induction curve. Before analysis or feature computation, the ChlF induction data were normalized by using the *Min-Max* method.

Statistical analysis was performed on the induction features with the statistical software package *SPSS* (*SPSS Inc.*, version 21.0, Chicago, USA). Statistical differences between different levels of drought stress were determined by using the least significant difference (LSD) method at the 0.05 probability level.

SVM, KNN, and the Ensemble toolbox in *MATLAB* (*The MathWorks*, Natick, MA, USA) were used to classify the plant samples into the three drought levels. The parameters or settings for each method were determined experimentally for the best classification accuracy. The cubic kernel was used as the SVM kernel function, the kernel scale was set to ‘automatic’, ‘boxconstraint’ level to 1, and the multiclass method to ‘one-vs-one’. The distance metric of KNN was set as ‘cosine’, the number of neighbors was set as 10, the distance weight was set as ‘equal’. The Ensemble method was set as ‘subspace’, learner type was set as ‘discriminant’, the number of learners was set as 30, and the subspace dimension was set as 13.

To test the generalization ability of the three methods, 80% of the samples were randomly selected as the training dataset, and the remaining 20% were selected as the test dataset.

Results

ChlF induction curves under three different drought stress durations were measured. It cannot be seen that drought stress resulted in differences in the curves due to ChlF variations between samples within groups (Fig. 1).

Induction features were computed and subjected to the least significant difference (LSD) analysis to determine if they are statistically different for different durations of drought stress treatments (Fig. 2). F_o , F_j , F_i , V_i , PI_{ABS} , ABS/RC , S_s , S_m , N , M_0 , DI_0/RC , and Area differed between the three drought levels. V_j , F_m/F_o , F_v/F_o , F_v/F_m , φE_0 , φP_0 , φD_0 , ψ_0 , and HACH Area do not show the

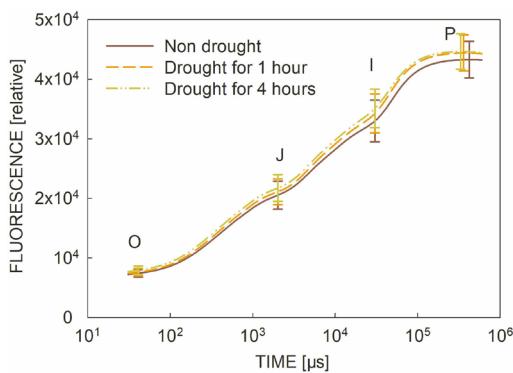


Fig. 1. Mean ChlF induction curves for three different drought levels (no drought, drought for 1 h, and drought for 4 h). The error bars are the standard deviations of O, J, I, and P points of the rice samples under the same drought stress level.

statistical difference between the non-drought (D0) and 1-h drought stress treatments (D1) while F_m , F_v , φPav , and ET_0/RC did not differ between the 1-h (D1) and 4-h (D4) drought stress treatments. Since the ChlF induction features varied, they are potentially useful for distinguishing drought levels in rice plants. It lays a base for the classification of different rice drought levels by using ChlF. However, the statistical difference does not mean that the classification accuracy for drought stress levels may be very high because of sample variations within groups.

Three classification techniques (SVM, KNN, and Ensemble) were applied to classify the plant samples into different drought levels with the commonly used maximum photochemical quantum yield of PSII in the dark (F_v/F_m), the induction features, or the whole induction curve shown in Fig. 2. Table 1 shows the maximum classification accuracy (Max), the minimum classification accuracy (Min), and the average classification accuracy (Average) of running the test dataset 10 times by SVM, KNN, and Ensemble, respectively. It can be seen that among the three classification methods, the average classification accuracy rate of the SVM method with the induction curve as input is the highest, which is 88.5%. In addition, the induction curve as input performed better than just the induction features except for the KNN method.

To test the consistency of classification by the three methods (SVM, KNN, and Ensemble) for drought levels of rice, the Cohen's kappa coefficient was calculated to analyze the consistency between the actual stress levels and the predicted stress levels. Generally, the kappa coefficient is between 0 and 1, which is divided into five intervals to represent the consistency between the actual category and the predicted category: 0.00–0.20 indicates very low consistency, 0.21–0.40 indicates fair consistency, 0.41–0.60 indicates moderate consistency, 0.61–0.80 indicates high consistency, and 0.81–1.00 indicates almost perfect consistency. The kappa coefficient values for the three classification methods are shown in Table 2. The kappa coefficient (0.82) of the SVM method with the induction curve as input is the highest. This result shows that SVM has great advantages in the qualitative analysis of rice drought levels.

For conciseness, only the results from SVM are further reported and discussed. The SVM classification results for the training dataset are shown in Fig. 3 and the classification results for the test dataset are listed in Table 4. Fig. 3 shows the confusion matrix of SVM classification of rice drought levels for the training dataset. The values in the confusion matrix represent the probability that the true drought stress levels are denoted by the vertical axis classified as the predicted drought stress levels denoted by the horizontal axis through the SVM classifier. The larger the diagonal value in the confusion matrix, the higher the classification accuracy. The total classification accuracy of drought stress levels is 47.0% (the mean of the truly positive results in Fig. 3A) only based on the maximum photochemical quantum yield of PSII (F_v/F_m) in Fig. 3A. Fig. 3B shows that the

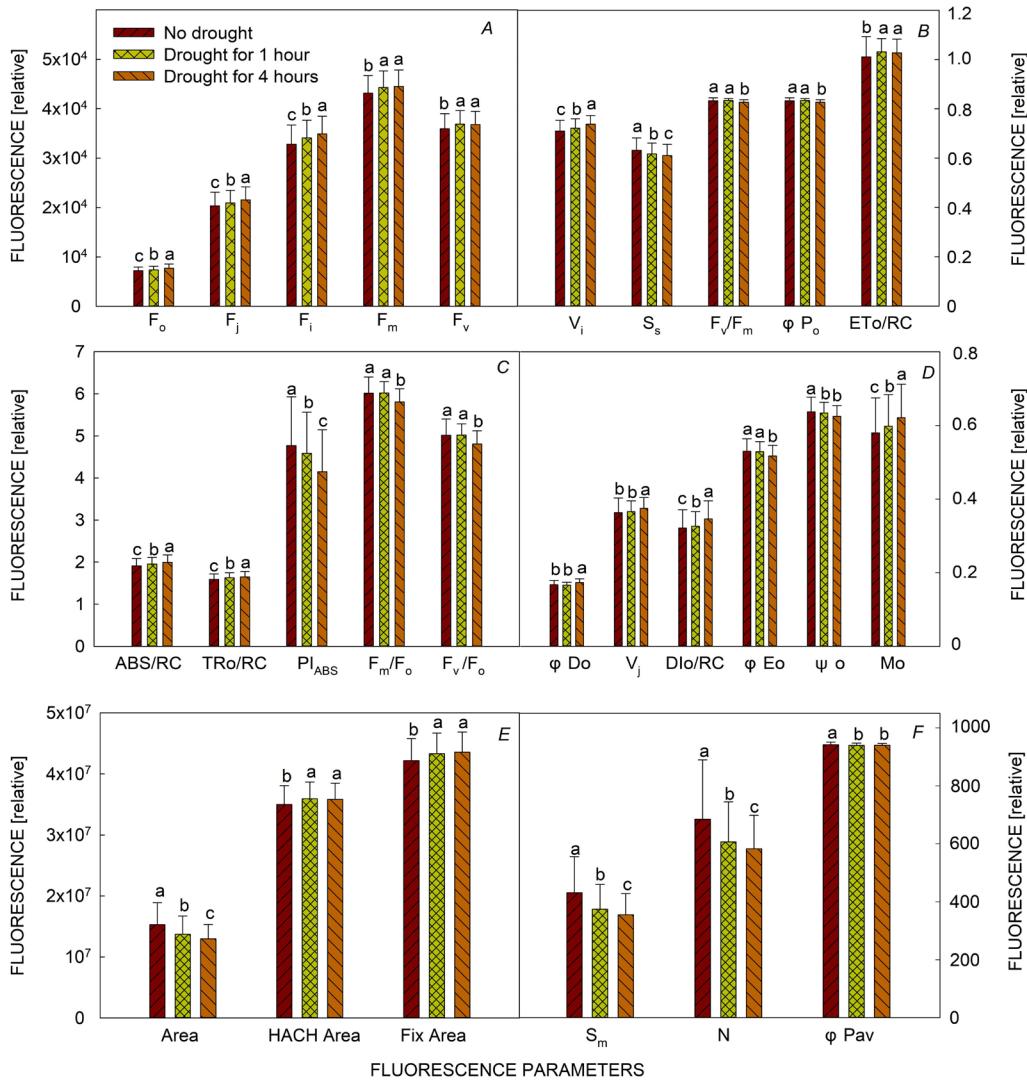


Fig. 2. One-way ANOVA of ChlF induction features under different drought levels (different lowercase letters indicate significant differences at $p < 0.05$). F_o – minimal chlorophyll a fluorescence intensity in the dark; F_j – chlorophyll a fluorescence intensity at the J step; F_i – chlorophyll a fluorescence intensity at the I step; F_m – maximal chlorophyll a fluorescence intensity; F_v – variable fluorescence intensity in the dark; V_j – relative variable fluorescence intensity at the J step; V_i – relative variable fluorescence intensity at the I step; F_m/F_o – electron transport through PSII; F_v/F_o – quantum efficiency of PSII; F_v/F_m – maximum photochemical quantum yield of PSII in the dark; M_0 – approximated initial slope (in ms^{-1}) of the fluorescence transient; Area – area between fluorescence curve and F_m (background subtracted); Fix Area – area below the fluorescence curve between $F_{40\mu\text{s}}$ and $F_{1\text{s}}$ (background subtracted); S_m – normalized area between ChlF induction curve and the line $F = F_m$, (multiple turnover); S_s – the smallest S_m turnover (single turnover); N – reduction times of Q_A from F_o to F_m ; ϕP_0 – maximum quantum yield of PSII; ψ_o – probability that a trapped exciton moves an electron further than Q_A^- ; ϕE_0 – quantum yield of electron transport; ϕD_0 – quantum yield of energy dissipation; ϕPav – average (from time 0 to t_{Pav}) quantum yield for primary photochemistry; PI_{ABS} – performance index for energy conservation from exciton to the reduction of intersystem electron acceptors; ABS/RC – absorption per reaction center; TR_o/RC – trapped energy flux per reaction center; ET_o/RC – electron transport per reaction center; DI_o/RC – dissipation per reaction center (at $t = 0$).

total classification accuracy of drought stress levels can reach 73.3% (the total classification accuracy of the three drought levels based on the induction features, which is the mean of the true positive rate in Fig. 3B) and the correct classification rate for the 4-h drought stress treatment (D4) can reach 80.0% when only induction features were used as the SVM model input. However, the classification accuracy can reach 88.3% [the total

classification accuracy of the three drought levels based on the induction curves, which is the mean of the truly positive rate in Fig. 3C and the classification rate for the 4-h drought treatment (D4) can reach 91.0% when the data of the whole induction curve was used as the SVM model input (Fig. 3C)]. Most of the misclassified cases were classified into the neighbor groups. For example, samples of D4 have an 80% chance to be classified as drought

Table 1. Comparison of drought stress level classification accuracy [%] obtained from different methods (SVM, KNN, and Ensemble) and inputs for the test dataset. F_v/F_m – maximum photochemical quantum yield of PSII.

Methods	SVM			KNN			Ensemble		
	Max	Min	Average	Max	Min	Average	Max	Min	Average
F_v/F_m [%]	46.24	43.59	45.45	45.13	38.46	41.95	44.62	40.51	41.91
Induction features [%]	75.21	72.14	73.21	68.21	61.71	65.26	68.03	63.76	65.88
Induction curve [%]	88.51	85.50	86.84	67.80	60.45	63.22	87.57	83.05	85.37

Table 2. Comparison of the kappa coefficients obtained from different methods (SVM, KNN, and Ensemble) for the test dataset. F_v/F_m – maximum photochemical quantum yield of PSII.

Methods	SVM			KNN			Ensemble		
	Max	Min	Average	Max	Min	Average	Max	Min	Average
F_v/F_m	0.23	0.15	0.18	0.18	0.08	0.13	0.17	0.11	0.13
Induction features	0.63	0.58	0.60	0.52	0.43	0.48	0.52	0.46	0.49
Induction curve	0.83	0.78	0.82	0.52	0.41	0.45	0.81	0.75	0.78



Fig. 3. Confusion matrix of Support Vector Machine (SVM) classification of rice drought levels for the training dataset. (A) Based on the maximum photochemical quantum yield of PSII (F_v/F_m), (B) based on induction features, (C) based on the induction curve. D0, D1, and D4 indicate no drought treatment, drought treatment for 1 h, and drought treatment for 4 h, respectively.

treatment for 1-h (D1) but unlikely as non-drought rice (D0) in Fig. 3C.

Tables 3, 4, and 5 show the SVM classification results for the test dataset based on the F_v/F_m , induction features, and induction curve, respectively. Table 3 shows the classification accuracy for the total rice samples is 43.9% based on F_v/F_m and the kappa coefficient value is 0.159. The classification accuracy for the total rice samples is 72.7% based on induction features as shown in Table 4 and the classification accuracy for the total rice samples is 86.7% based on induction curves as shown in Table 5. The total classification accuracy using the induction curves is 42.7% and 14.0% higher than that using F_v/F_m and the induction features, respectively. In general, using the induction curve as input proved to be advantageous over using only F_v/F_m or induction features as shown by the higher each or total correct classification percentages and a greater kappa coefficient value (0.80 vs. 0.60 or 0.16). The correct classification rates using the induction curve for the non-drought treatment and 4-h drought treatment were relatively high in Table 5, which should be related to the physiological changes of rice samples induced by drought stress.

Discussion

The difference of ChlF induction under different drought durations as shown in Fig. 1 and the statistical difference of some parameters under different drought durations as shown in Fig. 2 are consistent with findings in the literature. Under drought stress, different parts of electron transport of PSII are affected (Oukarroum *et al.* 2007, Zivcak *et al.* 2013, Bano *et al.* 2021). The changes of plant photosynthesis with external factors can be traced by measuring the ChlF induction and correlated with the drought stress (Saglam *et al.* 2014). The different steps and stages of ChlF induction can be linked to PSII, PSI, and the electron transfer efficiency between the two photosystems, components of photosynthetic electron transport

Table 3. Classification results for test dataset based on the maximum photochemical quantum yield of PSII (F_v/F_m). D0 – no drought stress treatment; D1 – drought stress treatment for 1 h; D4 – drought stress treatment for 4 h. Kappa coefficient = 0.16.

Actual samples	Predicted samples			Total
	D0	D1	D4	
D0	63	81	51	195
D1	48	87	60	195
D4	15	73	107	195
Total	126	241	218	585
Accuracy [%]	50.00	36.10	49.08	43.93

Table 4. Classification results for test dataset based on induction features. D0 – no drought stress treatment; D1 – drought stress treatment for 1 h; D4 – drought stress treatment for 4 h. Kappa coefficient = 0.59.

Actual samples	Predicted samples			Total
	D0	D1	D4	
D0	142	45	8	195
D1	37	128	30	195
D4	5	35	155	195
Total	184	208	193	585
Accuracy [%]	77.17	61.54	80.31	72.65

Table 5. Classification results for testing dataset of rice under different drought levels using induction curve. D0 – no drought stress treatment; D1 – drought stress treatment for 1 h; D4 – drought stress treatment for 4 h. Kappa coefficient = 0.80.

Actual samples	Predicted samples			Total
	D0	D1	D4	
D0	169	24	2	195
D1	13	169	13	195
D4	3	21	171	195
Total	185	214	186	585
Accuracy [%]	91.35	78.97	91.93	86.67

(Stirbet and Govindjee 2011, Brešić *et al.* 2015). Previous studies found that ChlF has a strong correlation with rice drought stress. In our study, we expanded the research by proposing a machine learning method based on ChlF to classify drought levels.

Our results show that there are statistical differences in some induction features under different drought durations, which is consistent with the results in the literature (Kumar *et al.* 2016). The classification accuracy of using all the data on the OJIP induction curve as inputs is higher than the accuracy of only using F_v/F_m or some ChlF feature parameters as inputs. In addition, the kappa coefficients in Table 5 also show that all the data on the OJIP induction curve as the model input lead to better classification performance for rice drought stress levels. This implies that drought stress level classification accuracy can be

significantly improved by using the whole ChlF induction curve although there is no clear visible difference between ChlF signals for rice samples with different levels of drought stress treatment. This also implies that there is rich useful hidden biological information in the ChlF induction curve and the information can be used in future research for more application through machine learning techniques.

SVM is a supervised learning method. Its decision boundary is the maximum margin plane of the learning sample solution. In addition, the regularization term is added to the solution system to optimize the structural risk, so it has certain advantages in classification. Therefore, compared with the KNN and the Ensemble model, the SVM model has better capability to classify different levels of rice drought stress. And the kappa coefficients in Table 4 also show that the consistency of the classification results using SVM is relatively better. Machine learning methods are useful for the analysis of ChlF induction curves, especially in rice drought classification, which has been proved in this paper. This work demonstrated SVM had advantages in classifying rice drought stress levels with the ChlF data and the whole OJIP induction contains more plant physiological information.

Conclusion: In this work, the SVM technique was used for the classification of drought stress levels based on ChlF. The results show that the SVM model is a useful tool in rice drought stress level classification. In addition, the whole OJIP induction contains more information about the physiological state of plants, which was often omitted in literature when only the ChlF characteristic features are used in the analysis. In future work, more ChlF data under different environmental conditions should be considered to extend the developed method. It is expected that this ideal will be useful to improve agricultural production and agricultural genetic breeding under environmental stress.

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Appendix. Chlorophyll *a* fluorescence parameters.

OJIP parameters	Definitions
F_o	Minimal chlorophyll <i>a</i> fluorescence intensity in the dark
F_j	Chlorophyll <i>a</i> fluorescence intensity at the J step
F_i	Chlorophyll <i>a</i> fluorescence intensity at the I step
F_m	Maximal chlorophyll <i>a</i> fluorescence intensity
$F_v = F_m - F_o$	Variable fluorescence intensity in the dark
$V_j = (F_j - F_o)/(F_m - F_o)$	Relative variable fluorescence intensity at the J step
$V_i = (F_i - F_o)/(F_m - F_o)$	Relative variable fluorescence intensity at the I step
F_m/F_o	Electron transport through photosystem II
$F_v/F_o = (F_m - F_o)/F_o$	Quantum efficiency of photosystem II
$F_v/F_m = (F_m - F_o)/F_m$	Maximum photochemical quantum yield of photosystem II in the dark
$M_0 = 4(F_{300} - F_o)/(F_m - F_o)$	Approximated initial slope (in ms^{-1}) of the fluorescence transient
Area	Area between fluorescence curve and F_m (background subtracted)
Fix Area	Area below the fluorescence curve between $F_{40\mu s}$ and F_{1s}
$S_m = Area/(F_m - F_o)$	Normalized area between ChlF induction curve and the line $F = F_m$
S_s	The smallest S_m turnover (single turnover)
$N = S_m \times M_0 \times (1/V_j)$	Reduction times of Q_A from F_o to F_m
$\varphi P_0 = 1 - (F_o/F_m)$ (or F_v/F_m)	Maximum quantum yield of PSII
$\psi_0 = 1 - V_j$	Probability that a trapped exciton moves an electron further than Q_A^-
$\varphi E_0 = (1 - F_o/F_m) \times (1 - V_j)$	Quantum yield of electron transport
$\varphi D_0 = 1 - \varphi P_0 = F_o/F_m$	Quantum yield of energy dissipation
$\varphi P_{av} = \varphi P_0 \times (1 - V_j)$	Average (from time 0 to t_{fm}) quantum yield for primary photochemistry
$PI_{ABS} = RC/ABS \times [\varphi P_0/(1 - \varphi P_0)] \times [\psi_0/(1 - \psi_0)]$	Performance index for energy conservation from exciton to the reduction of intersystem electron acceptors
$ABS/RC = (M_0/V_j)(1/\varphi P_0)$	Absorption per reaction center
$TR_o/RC = M_0 \times (1/V_j)$	Trapped energy flux per reaction center
$ET_o/RC = (M_0/V_j)(1 - V_j)$	Electron transport per reaction center
$DI_o/RC = ABS/RC - TR_o/RC$	Dissipation per reaction center (at $t = 0$)